



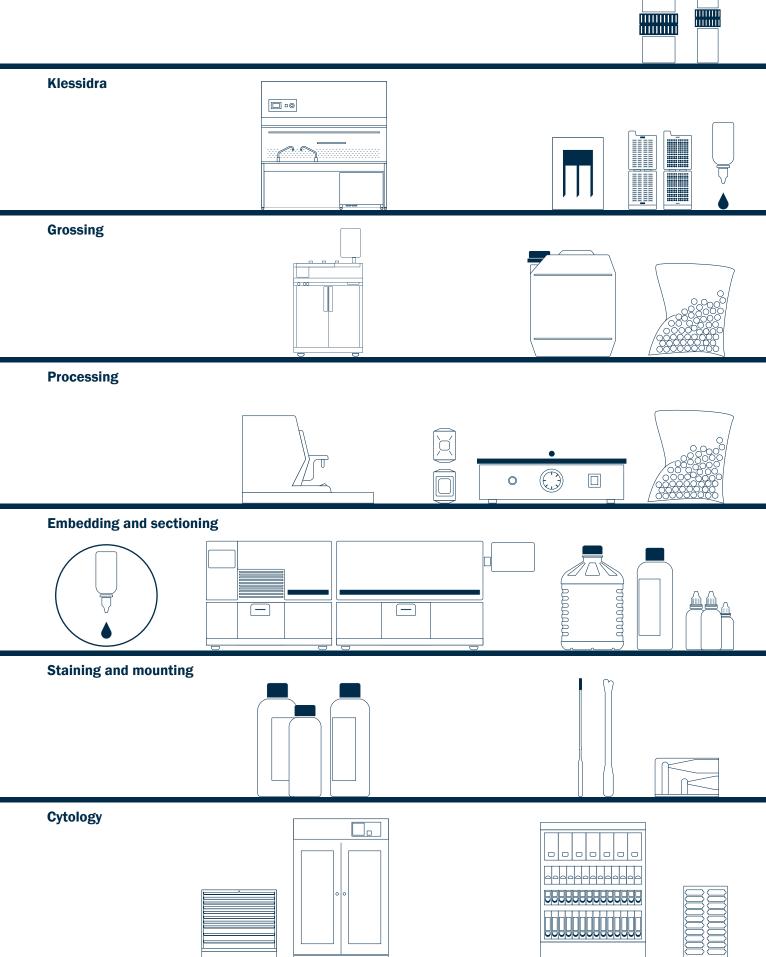
Bio - Optica

# **Improving Pathology**

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# **GENERAL CATALOGUE**

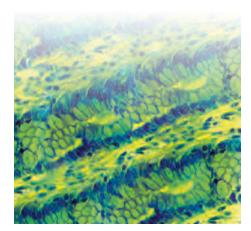




# HISTORY

Founded in **1977**, Bio-Optica, is a solid and established reality committed to anatomic pathology, and thanks to its complete and unique portfolio of instruments, reagents and consumables, it operates on the national and international markets.

# CERTIFIED QUALITY



The active collaboration with customers and a constant research for innovation, in order to reach high quality standards, is demonstrated by the **ISO 13485** certification and from May 2022 also **IVDR 2017/746 (CE)** compliant..

Reagents



Our mission is to provide the labs operators **the best way to do their job**.

We want to ensure the chance to make diagnosis **correctly, fast and avoiding mistakes**.

We want to help people having a long, healthy and good life.



**Consumables** 

# VISION

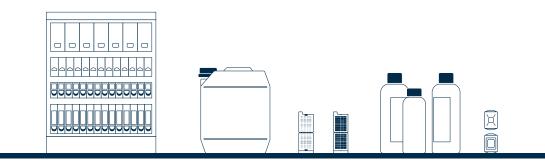
To create a network of **trusted distributors** all over the world and allow them to achieve more.

Be capable to understand **customer needs**, provide high standards and innovative solutions.

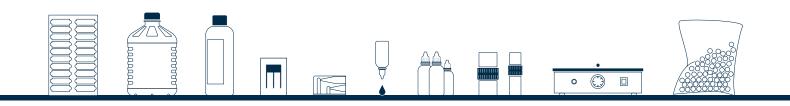
To build the most **dynamic business** capable of attracting every opportunity.



# Equipments







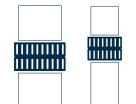
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Klessidra





Solution A: 30% buffered neutral formalin Solution B: phosphate buffer

## Klessidra

Klessidra is a patented, closed-circuit safety device, which prevents contact between formaldehyde and the user, in accordance with Commission Regulation (EU) No. 605/2014. It is intended for the fixation and transport of small histological samples. The device is provided by a mechanism which prevents the reflux of formalin into the previous container in order to avoid

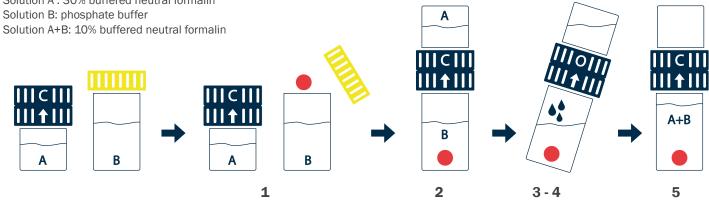
Ipofixation problems and the loss of the biopsies.

#### **Klessidra 30**

Klessidra 30 is supplied in pre-filled containers, the buffer solution helps operators remove biopsies from needles.

The container with two blue caps contains concentrated formalin. The buffered neutral 10% formalin reconstitutes after blending with the buffer solution.

	PRODUCT	PACK	CODE
•	Klessidra 30 10 ml formaldhevde 12% and 20 ml buffer	27 pcs.	05-01V15PKF
		07	
•	Klessidra 30 Blue 10 ml blue formaldheyde 12% and 20 ml buffer	27 pcs.	05-01V15PKFC
	Rack	2 pcs.	05-900900
•	Test tube rack for 16 Klessidras, to facilitate	2 μυ3.	03-900900
	transport within the hospital		
•	Klessidra transport		
	Safety box complete with foam with 21 places	1 pc.	05-900700
	Foam with 21 places	1 pc.	05-900800













### Klessidra 90

Klessidra 90 is the largest size in the Klessidra range. The 90 ml size can hold up to 12 Bio Cassettes. The maxi 160 ml version can hold two SuperMegaCassettes or 20 Bio Cassettes.

	PRODUCT	PACK	CODE
•	Klessidra 90 90 ml of 10% buffered neutral formalin	8 pcs.	05-01V125PK
•	Klessidra 160	8 pcs.	05-01V250PK
	160 ml of 10% buffered neutral formalin		

#### Instruction for use

- 1) Put the specimen into the empty container (B) with or without biocassette;
- 2) On a flat surface, connect the containers (solution A on top) and apply a slight pressure from above (red arrow) in order to get the correct alignment;
- 3) On a flat surface, screw the prefilled formalin container(A)on container(B) which contains the specimen;
- 4)Rotate the two lids on "open" position (until the arrow is aligned with the"O") and tilt the device to let the formalin flow in the lower container;
- 5) Rotate the two lids again on "close" position (until the arrow is aligned with the "C").













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#### **Trimming Tech histology hood**

Trimming Tech hoods are designed to the highest quality standards so as to meet all operator requirements in relation to the prevention of chemical risk during grossing of histological samples. Made of stainless steel, they are equipped with a multiple extractor system that extracts fumes from the work surface, from the front and from above. The control panel with soft-touch keypad provides an intuitive interface for setting the desired operating parameters.

#### **Construction features**

- Stainless steel structure
- Power-operated vertically sliding front safety glass sash for fume containment
- Filter basket and lid for the formalin container

#### Work surface features

- Non-drip lip
- Sink with pedal-operated mixer tap and pull-out shower head for washing the work surface
- Formalin disposal tank
- Waste fluid collection tanks with extractor and independent washing system

#### Extractor system

- Pre-installed alumina filters for formalin
- Cartridge-type synthetic fiber pre-filters pre-installed





## Grossing

<ul> <li>Trimming Tech 130 with sink on left 1300x750x2230 mm 50-130-001 with sink on right 50-130-002</li> <li>Trimming Tech 150 with sink on left 1500x750x2230 mm 50-150-001 with sink on right 50-150-002</li> <li>Trimming Tech 180 with sink on left 1800x750x2230 mm 50-180-001 with sink on right 50-180-002 with central sink 50-180-003</li> <li>Trimming Tech 180 with sink on the left and waste bin with lid on the right side with sink on the right and waste bin with lid on the left side</li> </ul>		PRODUCT	WORK SURFACE	DIMENSIONS	CODE
Trimming Tech 150       with sink on left       1500x750x2230 mm       50-150-001         with sink on right       50-150-002         Trimming Tech 180       with sink on left       1800x750x2230 mm       50-180-001         with sink on right       50-180-002         with central sink       50-180-003         Trimming Tech 180       with sink on the left and       1800x750x2230 mm       50-180-004         with waste bin       with sink on the left and       1800x750x2230 mm       50-180-004         with waste bin       with sink on the right and       50-180-005         with sink on the right and       50-180-005	•	Trimming Tech 130	with sink on left	1300x750x2230 mm	50-130-001
• With sink on right       50-150-002         • Trimming Tech 180       with sink on left       1800x750x2230 mm       50-180-001         • With sink on right       50-180-002       with central sink       50-180-003         • Trimming Tech 180       with sink on the left and       1800x750x2230 mm       50-180-003         • Trimming Tech 180       with sink on the left and       1800x750x2230 mm       50-180-004         with waste bin       waste bin with lid on the       right side       with sink on the right and       50-180-005         waste bin with lid on the       sate bin with lid on the       50-180-005       So-180-005			with sink on right		50-130-002
Trimming Tech 180       with sink on left       1800x750x2230 mm       50-180-001         with sink on right       50-180-002         with central sink       50-180-003         Trimming Tech 180       with sink on the left and       1800x750x2230 mm         with waste bin       with sink on the left and       1800x750x2230 mm         with waste bin       with sink on the right and       50-180-004         with waste bin       with sink on the right and       50-180-005         with sink on the right and       50-180-005	٠	Trimming Tech 150	with sink on left	1500x750x2230 mm	50-150-001
with sink on right       50-180-002         with central sink       50-180-003         Trimming Tech 180       with sink on the left and 1800x750x2230 mm 50-180-004         with waste bin       vaste bin with lid on the right side         with sink on the right and saste bin with lid on the right side       50-180-005			with sink on right		50-150-002
with central sink       50-180-003         Trimming Tech 180       with sink on the left and 1800x750x2230 mm 50-180-004         with waste bin       right side         with sink on the right and waste bin with lid on the right side       50-180-005         with sink on the right and waste bin with lid on the       50-180-005	٠	Trimming Tech 180	with sink on left	1800x750x2230 mm	50-180-001
Trimming Tech 180     with sink on the left and 1800x750x2230 mm 50-180-004     waste bin with lid on the     right side     with sink on the right and 50-180-005     waste bin with lid on the			with sink on right		50-180-002
with waste bin     waste bin with lid on the right side       with sink on the right and     50-180-005       waste bin with lid on the     waste bin with lid on the			with central sink		50-180-003
right side with sink on the right and 50-180-005 waste bin with lid on the	•	Trimming Tech 180	with sink on the left and	1800x750x2230 mm	50-180-004
with sink on the right and50-180-005waste bin with lid on the		with waste bin	waste bin with lid on the		
waste bin with lid on the			right side		
			with sink on the right and		50-180-005
left side			waste bin with lid on the		
			left side		

#### Accessories for grossing hoods

PRODUCT	CODE
Cutting board 35 x 45 cm	50-500-050
Millimeter ruler	50-500-054
Garbage disposal unit with pedal control (*)	50-500-055
UV lamp with self-switching off programming and rolling protection curtain. (*)	50-500-057
Formalin suction filter	50-500-058
Stainless steel filter for formalin sink	50-500-059
Magnetic knife-holder (*)	50-500-060
Paper handkerchief distributor	50-500-061
Stainless steel filter for water sink	50-500-062
Magnifying glass with neon light	50-500-069
UV lamp replacement	50-500-070
Cleaning kit	50-500-071
HEPA Filter (High Efficiency Particulate Air)	50-F005
Alumina filter for formalin	50-F017
Synthetic fiber pre-filter	50-F007
Swivel stool	40-300-450
Footrest	40-300-451
Waste trolley	50-500-075
(*) Accessories which can only be installed at the time of production	

 $(\ensuremath{^*})$  Accessories which can only be installed at the time of production









### **Protective equipment for laboratory work**

#### **Transport** bags

Polypropylene bags for transporting specimens, complete with grip seal closure and document folder.

DIMENSIONS	PACK	CODE
16 x 25 cm	500 pcs.	44-9590

#### **Bio-Pads**

Pads made of special fabric for absorbing formaldehyde. Ideal for containing spillages, leaks and drips of formaldehyde when grossing anatomical specimens, thus reducing the risk of operator exposure to formalin.

DIMENSIONS	PACK	CODE
203 x 254 mm	25 pcs.	08-FNP0810







#### **Accessories**

#### Endokit

Endokit is a complete patented system for the correct orientation of endoscopic biopsies, consisting of:

- Pre-cut strips of nitrocellulose with one slanted end
- Test tubes pre-filled with buffered neutral 10% formalin for immediate fixation of biopsies

PACK	CODE
40 Endokit strips and 80 pre-filled test tubes 40 Endokit	08-8700N 08-8710N



### **Grossing board**

Grossing boards for the dissection of anatomical specimens. The single-use grossing boards are equipped with a millimeter scale.

DIMENSIONS	MATERIAL	PACK	CODE
15 x 21 cm	Cardboard	20 pcs.	08-8000 (single-use)
30 x 21 cm	Cardboard	20 pcs.	08-8010 (single-use)
30 x 42 cm	Cardboard	20 pcs.	08-8020 (single-use)





#### **Bio Marking Dyes**

Bio-Optica marking dyes are special, non-toxic inks, made with natural polymers, used to mark surgical margins.

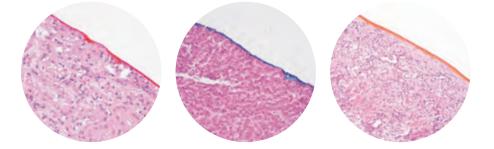
The advantages of using Bio Marking Dyes are as follows:

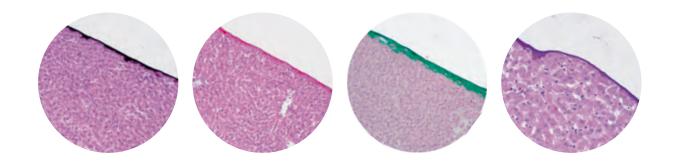
- They are non-toxic and made with natural polymers
- They dry in 2-3 minutes
- They do not require any additional work phases in other solutions to fix the color
- They do not spread into the tissue
- They can be applied to fresh or fixed samples
- They do not release color into solutions during fixing and processing











## 30 ml bottles with dispenser

COLOR	PACK	CODE
Blue	1 pc.	05-014-030-1
Black	1 pc.	05-015-030-1
Green	1 pc.	05-016-030-1
Yellow	1 pc.	05-017-030-1
Orange	1 pc.	05-018-030-1
Red	1 pc.	05-019-030-1
Violet	1 pc.	05-020-030-1

## 240 ml bottles

COLOR	PACK	CODE
Blue	1 pc.	05-014-240
Black	1 pc.	05-015-240
Green	1 pc.	05-016-240
Yellow	1 pc.	05-017-240
Orange	1 pc.	05-018-240
Red	1 pc.	05-019-240
Violet	1 pc.	05-020-240





#### **Multi-purpose containers**

Impact-resistant containers for the storage of small histological samples.

CAPACITY	PACK	CAP	CODE	
40 ml	500 pcs.	Screw	07-M40	
60 ml	500 pcs.	Screw	07-M60	

Multi-purpose containers with hermetically sealed press cap for the storage of histological samples, serigraphed with hazard symbols and risk phrases for formalin.

CAPACITY	PACK	CAP	CODE	
125 ml	250 pcs.	Press	07-7700	
250 ml	200 pcs.	Press	07-7750	
500 ml	100 pcs.	Press	07-7710	
1000 ml	100 pcs.	Press	07-7720	
3000 ml	50 pcs.	Press	07-7730	
5000 ml	20 pcs.	Press	07-7740	

#### Buffered neutral 10% formalin in pre-filled containers

CAPACITY	PACK	CAP	CODE
10 ml	80x5 ml	Screw	05-01P05
35 ml 55 ml	54x9 ml 54x18 ml	Screw Screw	05-01V15P 05-01V30P
55 ml 125 ml	54x28 ml 16x75 ml	Screw Screw	05-01V60P 05-01V125P16
250 ml	16x130 ml 6x300 ml	Press	05-01V250P16 05-01V500P
500 ml 1000 ml	6x600 ml	Press Press	05-01V1000P
3000 ml 5000 ml	4x1.800 ml 4x3.000 ml	Press Press	05-01V3000P 05-01V5000P

#### **Ready-to-use formalin**

	PRODUCT	PACK	CODE
•	Buffered neutral 10% formalin	4x2.5 I	05-01005Q
		1x5 I	05-01004F
		1x10	05-K01009
		1x20 I	05-K01004
•	Tap for tanks (10 and 20 I)	1 pc	05-1348





## Grossing

#### **Concentrated formalin**

	PRODUCT	PACK	CODE
•	38-40% formaldehyde	4x 2.5 I	05-01007Q
		1x20 I	05-K01007
٠	Concentrated buffered neutral formalin	1x10	05-K01004/CO

#### **Other fixatives**

	PRODUCT AND DESCRIPTION	PACK	CODE
٠	Bouin	1x500 ml	05-M01008
	For bone marrow biopsies	4x2.5 l	05-01008Q
•	Carnoy	1x2.5	05-01013E
	Product of choice for glycogen		
•	Hollande	1x2.5	05-01030E
	Excellent for trichrome staining		

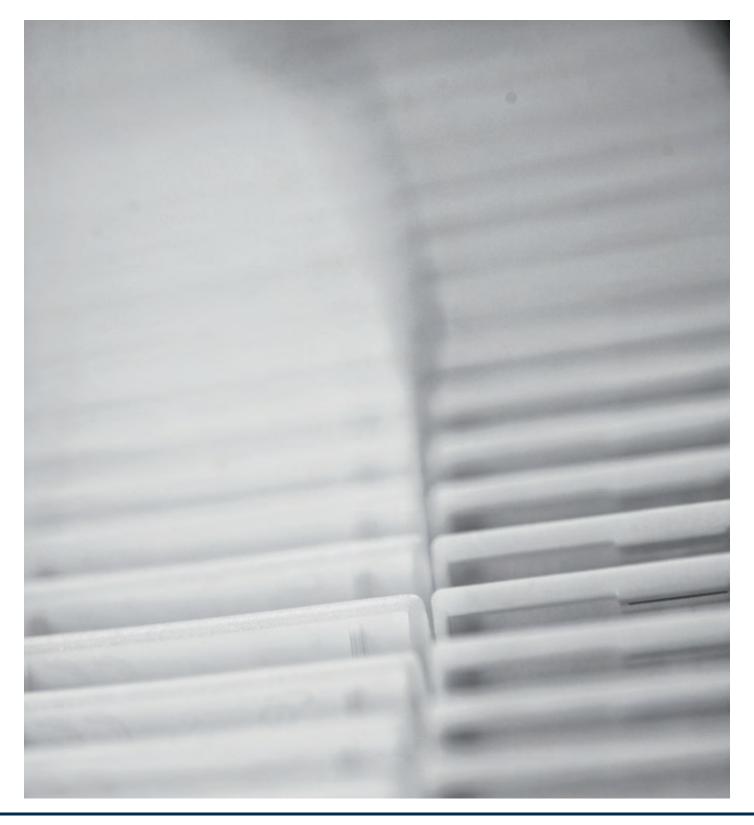
## **Decalcifying agents**

Decalcifying and/or fixative solutions for bone marrow biopsies and calcified tissues.

	PRODUCT AND DESCRIPTION	PACK	CODE
•	Osteodec	1x500 ml	05-M03005
	Decalcifying agent for bone biopsies	4x2.5 I	05-03005Q
٠	Biodec R	1x500 ml	05-M03009
	Rapid decalcifying agent for mineralized tissue	4x2.5 l	05-03009Q
٠	Electrolytic decalcifying agent	1x500 ml	05-M03004
	Decalcifying mixture comprising formic acid and hydrochloric acid	4x2.5 l	05-03004Q







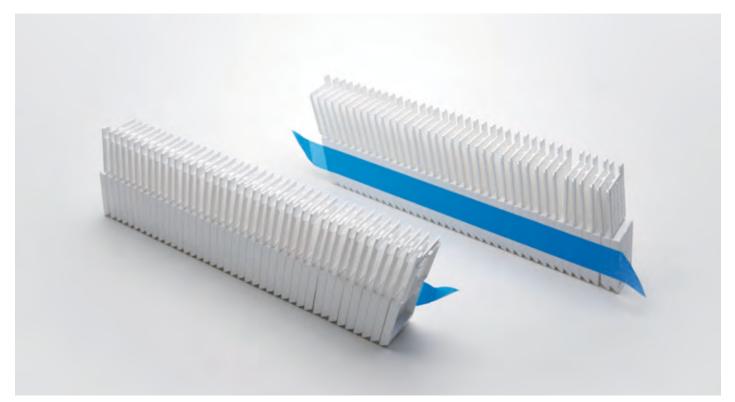


Grossing

## Stacked Bio Cassettes with lid with tape for printers

Bio Cassettes in polyacetal resin compatible with the main printing systems available on the market.

COLOR	PACK	CODE
White	40 stacks of 40 pcs.	07-9700
Orange	40 stacks of 40 pcs.	07-9710
Blue	40 stacks of 40 pcs.	07-9720
Yellow	40 stacks of 40 pcs.	07-9730
Lilac	40 stacks of 40 pcs.	07-9740
Pink	40 stacks of 40 pcs.	07-9750
Green	40 stacks of 40 pcs.	07-9760



#### COMPATIBILITY WITH COMMERCIALLY AVAILABLE WRITING SYSTEMS

CODE	PRIMERA	LEICA IPC	SAKURA SMART WRITE	SAKURA AUTO WRITE	THERMO PRINT MATE	HANDWRITING
07-9700	V	V	V	V	х	х
07-9710	V	V	V	V	Х	Х
07-9720	V	V	V	V	Х	Х
07-9730	V	V	V	V	Х	Х
07-9740	V	V	V	V	Х	Х
07-9750	V	V	V	V	Х	Х
07-9760	V	V	V	V	Х	Х
-						











### **Bio Cassettes**

• Bio Cassettes (with lid) Made of acetal resin for embedding standard samples.

COLOR	PACK	CODE
White	3x500 pcs.	07-7100
Orange	3x500 pcs.	07-7110
Blue	3x500 pcs.	07-7120
Yellow	3x500 pcs.	07-7130
Lilac	3x500 pcs.	07-7140
Pink	3x500 pcs.	07-7150
Green	3x500 pcs.	07-7160
Gray	3x500 pcs.	07-7180

#### • Bio Cassettes II (with separate lid)

COLOR	PACK LIDS + CASSETTES	CODE
White	1x2000 pcs. + 2x1000 pcs.	07-8100
Orange	1x2000 pcs. + 2x1000 pcs.	07-8110
Blue	1x2000 pcs. + 2x1000 pcs.	07-8120
Yellow	1x2000 pcs. + 2x1000 pcs.	07-8130
Pink	1x2000 pcs. + 2x1000 pcs.	07-8150
Green	1x2000 pcs. + 2x1000 pcs.	07-8160

• **Biopsy Cassettes (with lid)** Acetal resin cassettes for embedding biopsies and small samples.

COLOR	PACK	CODE
White	3x500 pcs.	07-7200
Orange	3x500 pcs.	07-7210
Blue	3x500 pcs.	07-7220
Yellow	3x500 pcs.	07-7230
Pink	3x500 pcs.	07-7250
Green	3x500 pcs.	07-7260
Gray	3x500 pcs.	07-7280

#### • Biopsy Cassettes II (with separate lid)

COLOR	PACK LIDS + CASSETTES	CODE
White	1x2000 pcs. + 2x1000 pcs.	07-8200



## Grossing

## Mega Cassettes (double height, with lid)

COLOR	PACK	CODE
White	750 pcs.	07-7300

#### Super Mega Cassettes (for large samples with separate lid)

Designed with larger mesh to increase the contact surface of the paraffin and prevent the sample from detaching during microtome cutting.

COLOR	PACK	DIMENSIONS	CODE
White	200 pcs.	70x50x15 mm	07-7000

## **Embedding Cassettes**

Acetal resin cassettes with round holes, for use with metal lids.

DESCRIPTION	PACK	CODE
White cassettes	3x1.000 pcs.	07-7350
Metal lid	10 pcs.	07-086-74195

#### **Biopsy pads and Histoshield**

Foam pads made of special material that is permeable to solvents and paraffins. They facilitate the processing of very small samples without any risk of loss of material.

Histoshield for laser printers	500 pcs.	07-7292
Biopsy pads Black	5000 pcs.	07-7291
Biopsy pads Blue	500 pcs.	07-7290
PRODUCT AND COLOR	PACK	CODE
	P. 0./	

#### **Biopsy bags**

Fine-mesh polyester bags, resistant to paraffin and solvents. Ideal for processing small histological samples.

DIMENSIONS	PACK	CODE
30x45 mm 45x60 mm	1000 pcs. 1000 pcs.	07-00005 07-00003















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Processing

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### B-PR0450 – Automatic vacuum tissue processor

#beprofessional

The B-PRO450 is a floor-standing automatic vacuum system designed for routine processing of histological samples.

**15 Tanks System:** This means that you don't have to reduce steps in your conventional protocols. Using a single protocol for all types of samples, you can save time and improve the quality of results.

**B-PROtocols:** certified tested protocols, providing high level of standardization (IVDR compliant).

**#B-Support:** be always connected to the technical assistance department: real-time remote support.

B-CHECK: initial self-diagnosis that prevents possible failures.

**Process reliability:** integrated APC (Automated process completion) system as warranty of the process completion.

**Operator safety:** double filtration system and anti-burning technology (ABT): a solid wax discharge mechanism.

Easy to charge: 3 paraffin pre-melting elements.

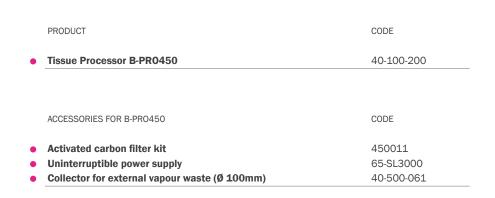
**RFID reagent traceability:** the processing is guaranteed by the unique traceability of the reagents using RFID technology.

**B-Friendly:** an integrated 15' touch screen with a simple and intuitive graphical user interface, making it easy to use and manage.

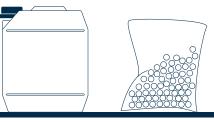
**Safe reagents loading system:** 3 optical sensors for the partial loading; 1 optical sensor to control the overflow.

#### Characteristics

Dimensions:	850 x 750 x 1650 mm (W x D x H)
Weight:	250 kg
Operating capacity:	Up to 450 cassettes per cycle
Number of storable protocols	20 programs (4 IVDR certified, 2 washing and 1 reverse)









#### **Pre-filled tanks**

PRODUCT	PACK	CODE
Formalin 10% buffered	5   x 1 tank	450001
Distilled water	5   x 1 tank	450003
B-Alcohol 70 per B-PRO 450	5   x 1 tank	450004
B-Alcohol 95 per B-PRO 450	5   x 1 tank	450005
B-Alcohol 100 per B-PRO 450	5   x 1 tank	450006
X-free	5   x 1 tank	450007
Xilene	5   x 1 tank	450009
Paraffin disposal tank	5   x 1 tank	450010
BioWax - Paraffin	3,8 kg x 3 bag	450012













#### Dehyol 70

A 70  $^\circ$  alcohol mixture that makes an ideal substitute for 70  $^\circ$  ethanol in all histology/ cytology procedures.

PACK	CODE
1x5 I	06-10075F
4x2.5 l	06-10075Q

#### Dehyol 95

A 95  $^\circ$  alcohol mixture that makes an ideal substitute for 95  $^\circ$  ethanol in all histology/ cytology procedures.

PACK	CODE
1x5 I	06-10070F
4x2,5 l	06-10070Q

### **Dehyol Absolute**

An absolute alcohol mix that makes an ideal substitute for absolute ethanol in all histology/cytology procedures.

PACK	CODE
1x5 I	06-10077F
4x2,5 I	06-10077Q

## AlcoolPath 95

A 95° alcohol mix comprising ethyl alcohol.

PACK	CODE
1x5	06-10031F
4x2,5 l	06-10031Q

#### **AlcoolPath Absolute**

An absolute alcohol mix comprising ethyl alcohol.

CODE
06-10030F
06-10030Q





Processing

#### Unyhol

Unyhol is an alcohol mix formulated to prevent excessive dehydration of samples and to substitute the full range of alcohol concentrations.

PACK	CODE
1x5	06-10071F
4x2,5 I	06-10071Q

#### **Bio Clear**

Clearing reagent of natural origin, formulated to replace xylene in the processing, de-waxing and dehydration of slides.

PACK	CODE
4x2.51	06-17820

#### Isopar Ultra

Clearing Isoparaffinic solvent, formulated to replace xylene in the processing, de-waxing and dehydration of slides.

PACK	CODE
1x5	06-1306F

#### **Xylene for histological applications**

Xylene-based solvent for histology and cytology procedures.

PACK	CODE
4x2,5 l	06-1304Q
1x5	06-1304F

#### X-Free

A xylene substitute solvent for use in histology and cytology procedures.

PACK	CODE
4x2.5 l	06-1305Q
5	06-1305F

#### **Bio-Wax Paraffin**

Mixture of pure paraffin (aliphatic hydrocarbons) and additive polymers (concentration: 0.3%) for histological samples embeddings. DMSO-free.

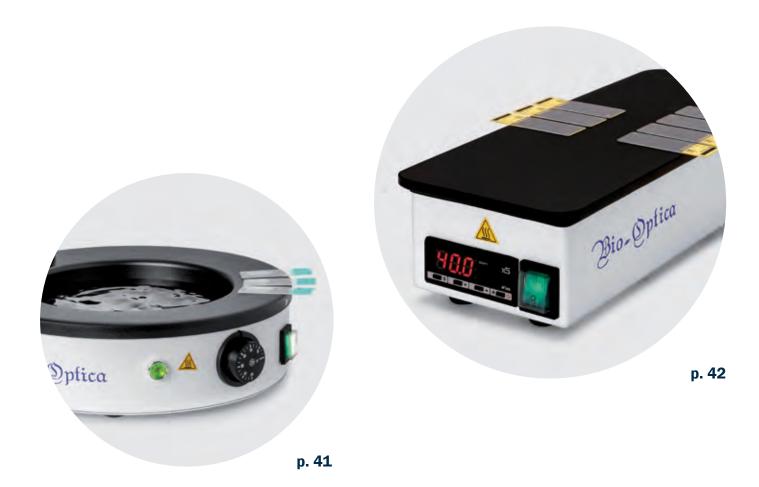
Bio-Wax Parafin	56÷58°C	8 bags x 1kg	08-7960
DESCRIPTION	MELTING POINT	PACK	CODE

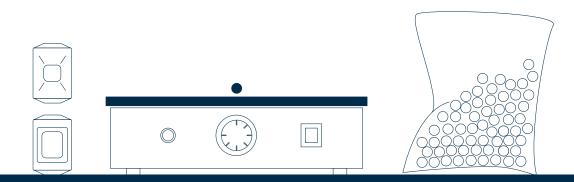












Embedding and sectioning



#### **Embedding station**

Modular specular system for embedding histological samples in paraffin. Comprises two separate units:

- Paraffin dispenser and thermal unit
- Cooling plate

The operating parameters, switch-on and switch-off of the two modules can be programmed separately.

The BEC150 paraffin dispenser is equipped with the following:

- Proximity sensor for dispensing paraffin, with flow control function
- 6 heated, pull-out holders for forceps
- 2 paraffin collection drawers with disposable containers
- Peltier spot capable of accommodating embedding molds for large samples
- Double thermal unit capable of accommodating the racks of any floor-standing processor
- Double jack for heated forceps or pestle
- Touch-screen monitor
- Fully lit work surface





## **Embedding and sectioning**

Accurate thermostatic control of the paraffin tank and work plate, and separate heating of the dispensing nozzle keep the operating temperature constant at all times.

The BCP170 cooling plate can accommodate up to 70 standard embedding cassettes and can be positioned on the right or the left of the paraffin dispenser according to the needs of the operator.

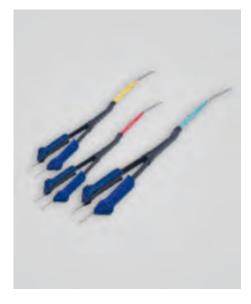
The dispenser can be ordered with the optional BCP230 cooling plate.













### **BEC150**

# • Characteristics

Overall dimensions:	560 x 605 x 405 mm (W x D x H)
Total weight: 18 kg	
Heated plate:	anodized aluminum, surface 517 x 120 mm (w x d)
Paraffin tank:	aluminum, volume 4 liters approx
Chambers (two) for processor aluminum, surface 225 x 160 mm (W x D)	
racks with removable tank:	
Peltier plate dimensions:	80 x 65 mm
Adjustment range:	heated elements: +20°C to +75°C
	Peltier plate: -3°C

# BEC170

#### • Characteristics

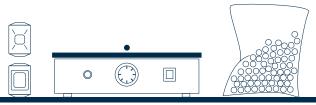
Overall dimensions:	410 x 605 x 405 mm (W x D x H)
Cooling surface dim.	370 x 350 mm (W x D)
Total weight:	24 kg
Temperature:	Working Temperature fixed at -10 °C

# **BEC230**

•

# • Characteristics

Overall dimensions:	410 x 605 x 405 mm (W x D	x H)
Cooling surface dim.	370 x 350 mm (W x D)	
Total weight:	24 kg	
PRODUCT		CODE
BEC150 paraffin dispenser		40-200-002
BCP170 cooling plate		40-300-202
BCP230 cooling plate		40-300-203
ACCESSORIES		CODE
Heated forceps - size 1 mm	- red	40-200-062
Heated forceps smooth - spa	are forceps size 2 mm - yellow	40-200-063
Heated forceps serrated – sp	pare forceps size 4 mm - blue	40-200-064
		10 000 005
Heated Tamper – size 8x8 m		40-200-065
Heated Tamper – size 16x16		40-200-066
Heated Tamper – size 28x25	mm	40-200-067
Foot switch		40-200-060
Magnifier lens - (code 40-20)	0-068 required)	40-200-061
Support for Magnifying Lens		40-200-068
Wax scraper		40-200-070
Paraffin recovery tray		40-200-071



# **Embedding and sectioning**

# Steel embedding molds

Stainless steel molds for embedding histological samples in paraffin.

# Bio Mold

MOLD DIMENSIONS	PACK	CODE
7 x 7 x 5 mm	12 pcs.	07-BM775
15 x 15 x 5 mm	12 pcs.	07-BM15155
24 x 24 x 5 mm	12 pcs.	07-BM24245
30 x 24 x 5 mm	12 pcs.	07-BM30245
37 x 24 x 5 mm	12 pcs.	07-BM37245

# Mega Mold

Super Mega Mold		
33 x 24 x 12 mm	6 pcs.	07-MBM6
MOLD DIMENSIONS	РАСК	CODE

MOLD DIMENSIONS	РАСК	CODE
65 x 45 x 15 mm	10 pcs.	07-7010

# **PVC** dispomold

Single-use, PVC embedding molds.

MOLD DIMENSIONS	PACK	CODE
7 x 7 x 5 mm	1500 pcs.	07-MP7070
15 x 15 x 5 mm	1500 pcs.	07-MP1515
24 x 24 x 5 mm	1500 pcs.	07-MP2424
30 x 24 x 5 mm	1500 pcs.	07-MP3024
37 x 24 x 5 mm	1500 pcs.	07-MP3724





# BCP230 freezing plate

Cooling plate equipped with a work tray with 48 mm high edge, designed to provide a refrigerated chamber and not just a cold work surface. The chamber is equipped with a lid. The advantages of this solution are as follows:

- High cooling power
- No condensation on the work surface or dripping on the bench
- Ability to cool a larger number of paraffin-embedded blocks (up to 300 approx) thanks to the special guides, which are available to order.

#### • Characteristics

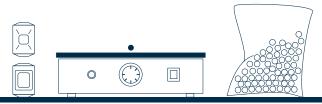
BCP230 freezing plate

Dimensions:	410 x 600 x 385 (L x W x H)	
Weight:	30 kg	
Capacity:	up to 300 cassettes approx	
Operating temperature:	to -20°C	
Cooling system:	CFC-free	
PRODUCT	CODE	

JODL

40-300-203





# **Embedding and sectioning**

#### WB1770 water bath

The WB1770 water bath is equipped with a removable Pyrex basin which is easy to fill with water and empty, and safer and more practical to use.

The temperature is controlled by a probe in direct contact with the water, which ensures absolute precision. It is equipped with a heated upper work surface capable of accommodating up to 24 slides.

#### Characteristics

Dimensions:	350 x 365 x 155 mm (L x W x H)	
Dimensions of slide-drying surface:	350 x 100 mm	
Weight:	8 kg	
Thermostat:	electronic thermostat with digital display	
Bath temperature:	+20°C to +70°C	
Temperature sensor:	NTC10K probe immersed directly in the water with	
	movable arm	
Basin lighting:	6 Watt neon lamp	
Slide plate temperature:	+20°C to +50°C	
PRODUCT	CODE	
WB1770 water bath	40-300-000	

#### WB100 round water bath

The WB100 round water bath is small, simple and reliable. It is equipped with an analog thermostat, heating indicator light and wide heated surface for 24 slides.

Dimensions mm 345x100 (ø x h) and dimensions of internal basin mm 225x50 (ø x h).

	PRODUCT	BATH TEMPERATURE	CODE
•	WB100 round water bath	+30 °C to +80 °C	40-300-002





### PC800 hot plate

The PC800 hot plate can dry 30 slides simultaneously. It is equipped with an anodized aluminum work surface and digital electronic thermostat. The temperature is adjustable up to 90 °C.

PRODUCT	DIMENSIONS	CODE
PC800 plate	150 x 380 x 100 mm	40-300-301



### **Brushes**

For microtomy and cryo-microtomy.

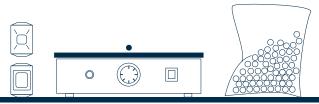
PRODUCT	PACK	CODE	
Small for collecting biopsies	4 pcs.	08-0822	
Small for collecting microtome sections	4 pcs.	08-0823	
Medium for collecting microtome sections	4 pcs.	08-0824	
Medium for microtome blade cleaning	2 pcs.	08-0825	
Large for microtome cleaning	2 pcs.	08-0826	
Large for cryostat cleaning	2 pcs.	08-0827	_
Set of 5 cryostat brushes	1 pc.	08-0828	
(Includes codes 0822-0823-0824-0825-0827)			
Set of 5 microtome brushes	1 pc.	08-0829	
(Includes codes 0822-0823-0824-0825-0826)			_



# **BioParaFree**

De-waxing solution in spray form, completely odorless, for cleaning paraffin residues from microtomes, embedding stations and work benches. Supplied in a bottle with nebulizer.

PACK	CODE
1x100 ml	08-1750



# **Embedding and sectioning**

#### Killik

Non-toxic embedding medium for the preparation of histological tissue for cryostat cutting.

COLOR	PACK	CODE
Neutral	4x100 ml	05-9801



# Crio Clor 0,3

Cryostat disinfectant spray comprising chlorhexidine (0.3%), effective against bacteria, fungi and other viruses (hepatitis B, poliomyelitis, herpes simplex).

1×125 ml	05.0800
1x125 ml	05-9802



#### **Cryo-Spray**

Histological freeze spray: for fast freezing of tissues for cryostat cutting, and for cooling paraffin-embedded samples before microtome cutting. The new formulation is non-flammable and contains no fluorinated gases. It is therefore safe for the environment and for operators.

PACK (

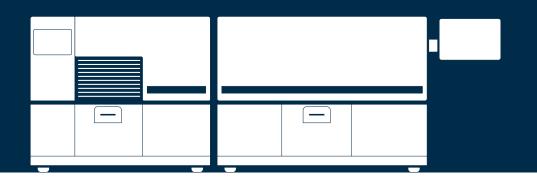
CODE

08-SPRAY

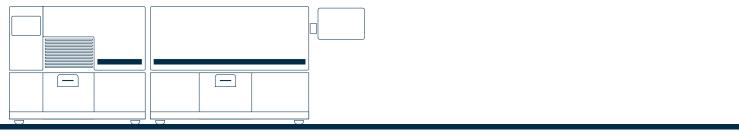












#### AUS240 PLUS Automatic Stainer

The AUS240 PLUS is an automated histology stainer with X-Y-axis transfer arm, fully programmable and suitable for all histo-cytological stains, both routine and special. It can perform multiple staining processes simultaneously. Their number is limited only by the number of dishes available (indicatively 10-12 processes).

Continuous loading of racks of 30 slides, with throughput dependent on the staining protocol. Equipped to impart a waving movement on the reagent dishes so as to reduce the quantity of precipitates in the dishes, thus keeping the reagent fresh at all times. The AUS240 can be integrated with the CVR 909, the automatic coverslipper.

#### Characteristics

Overall dimensions:	1220 x 780 x 770 mm (L x W x H)
Monitor:	+ 400 mm (I)
Total weight:	155 kg
Reagent work stations:	28
Water work stations:	5
Heated drying stations:	2 (60°C)
Unloading stations:	3
Loading stations:	2
Dish capacity:	485 ml
Slide rack:	capacity 30 slides
Number of programs:	up to 18 programs of over 100 steps each
Programmability:	each station can be set with an immersion time of 1" to
	99'59" (with tolerance of 1")
Interface:	large color touch-screen monitor for displaying the
	progress of work protocols, the layout of the process
	baths and all parameters relating to the staining cycles
	in progress
Safety:	Efficient, integrated fume filtration system

#### PRODUCT

CODE

AUS240 PLUS automatic stainer 40-400-350

# Stainings optimized for the instrument

PRODUCT	PACKAGING	CODE
Mayer hematoxylin	2,5	05-06002E
Eosin Y alcoholic solution 0,5%	2,5	05-10009E
Papanicolaou Harris hematoxylin	2,5	05-12011E
Papanicolaou EA50	2,5	05-12019E
Papanicolaou OG6	2,5	05-12013E
Giemsa	1500 tests	04-257000





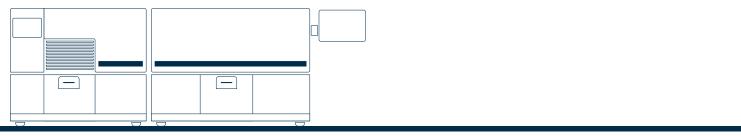
# Special Stainings optimized for the instrument

PRODUCT	PACKAGING	CODE
Alcian blue pH 2,5	1x1	04-160802/L
Alcian blue pH 2,5 PAS	1x1	04-163802/L
PAS	1x1	04-130802/L
Ziehl Neelsen Fite	1x1	04-111802/L

#### **CVR909 PLUS Automatic Coverslipper**

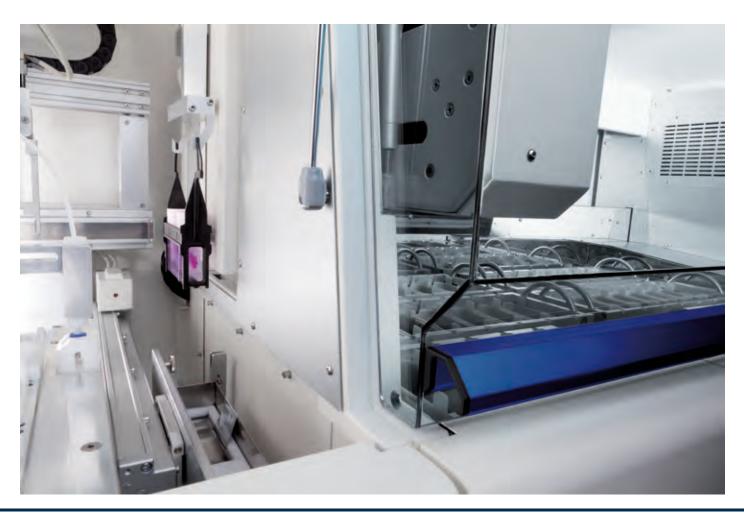
The CVR909 automatic coverslipper is the only instrument on the market that arranges the coverslipped slides directly on handy stackable trays, which are resistant to chemical reagents. The instrument is easy to use and specifically developed to facilitate routine laboratory operations. Continuous cleaning of the dispenser needle ensures high-quality coverslipping. The barcode reader ensures that all laboratory processes are fully traceable. Integrated with the AUS240 stainer, the coverslipper fully automates the process of de-waxing, staining and mounting of samples. It is possible to use three different sizes of coverglass (24x40 - 24x50 - 24x60) and to set the mounting medium quantity and dispensing mode.

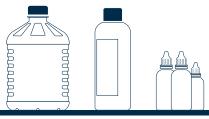




# Characteristics

Overall dimensions:	860 x 780 x 770 mm (L x W x H)
Total weight:	80 kg
Throughput:	180 slides/hour (directly on tray)
Mounting medium:	500 ml container
Output of mounted slides:	9 trays of 10 slides each (total: automated output of 90
	slides without operator intervention)
PRODUCT	CODE
CVR909 PLUS automatic coverslip	per 40-500-000







# Bench Tech benchtop fume hood

Laboratory fume hood with extraction from top and front, to be used for manual staining, slide mounting or as an area for liquid transfer. It is suitable for any laboratory benches.

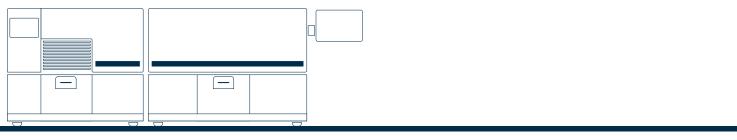
#### • Characteristics

	Extractor system:	1 three-phase spark-proof adjustat	ole electric extractor fan
	Lighting:	2 LED tubes, total 1500 lux	
	Control panel:	touch-screen monitor for the contr	ol and display of all
		functions	
	PRODUCT	DIMENSIONS	CODE
•	Benchtop fume hood 90	900 x 750 x 1340 mm	50-090-201
•	Benchtop fume hood 130	1300 x 750 x 1340 mm	50-130-201
٠	Benchtop fume hood 150	1500 x 750 x 1340 mm	50-150-201

### Filters

The filters are easy to change thanks to the handy Bio-Optica system with removable front panel. This system is used in all Bio-Optica fume hoods.

PRODUCT	CODE
HEPA filter	50-F005
UV lamp with auto switching off	50-500-057
Synthetic pre-filter	50-F007
Filter for solvents	50-F018
UV lamp replacement	50-500-070



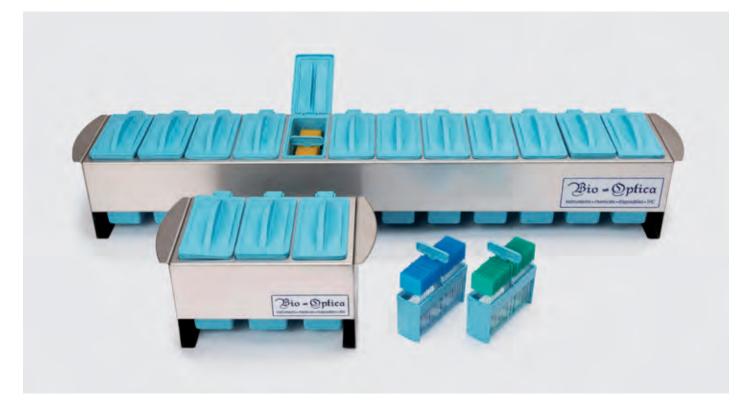
# Manual staining set

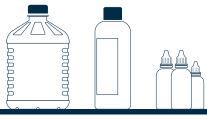
The simplest and most economical cyto-histological staining system, made of thermoplastic resin: the 10-10 manual staining set consists of twelve dishes with lid (capacity 300 ml or 80 ml) in a steel structure and a slide-rack for twenty-five slides. The hematology version consists of just three dishes with lid and one slide-rack. The dishes and rack are resistant to solvents and high temperatures (up to 170°C) and can be used in microwave ovens.

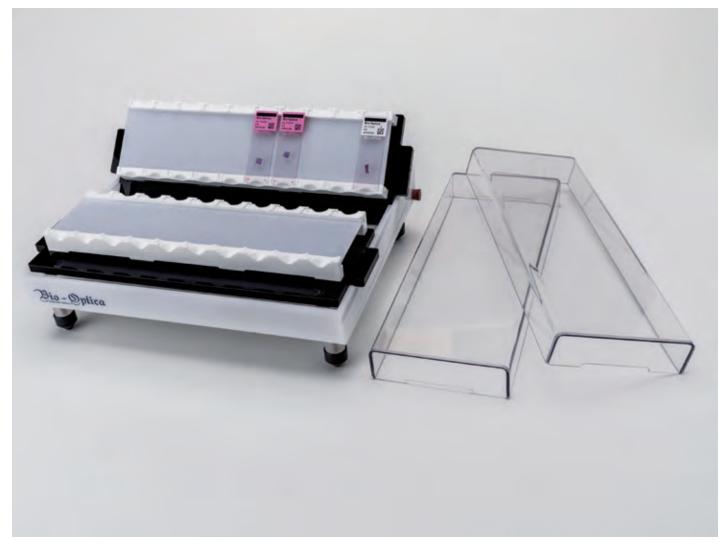
PRODUCTNo. OF DISHESCAPACITY of each dishCODEHematology set3300 ml10-20Hematology set380 ml10-21Staining set12300 ml10-10Staining set1280 ml10-11	_				
Hematology set3300 ml10-20Hematology set380 ml10-21		Staining set	12	80 ml	10-11
Hematology set 3 300 ml 10-20		Staining set	12	300 ml	10-10
		Hematology set	3	80 ml	10-21
PRODUCT No. OF DISHES CAPACITY of each dish CODE		Hematology set	3	300 ml	10-20
		PRODUCT	No. OF DISHES	CAPACITY of each dish	CODE

# PARTS AND ACCESSORIES

PRODUCT	No. OF DISHES	CAPACITY of each dish	CODE
Dish with lid attached	12 pcs.	300 ml	10-30
Dish with separate lid	12 pcs.	300 ml	10-33
Dish with separate lid	12 pcs	80 ml	10-34
Rack for 25 slides with plastic handle	6 pcs.		10-42
Steel slide basket for 8 slides	1 pc		10-44







#### Slide master for special and immunohistochemistry staining

Slide Master is the ideal manual stainer for special and immunohistochemistry staining. Equipped with 20 stations, humid chamber and lid, it allows horizontal incubation and inclined washing.

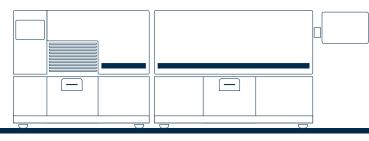
The adjustable feet and spirit level help keep the work surface totally horizontal.

DIMENSIONS

CODE

32 x 26 x 11 (L x W x H) cm

15-MEQ001





# Timer

Solvent-resistant laboratory timer.

MODEL	PACK	CODE
Digital electronic	1 pc.	44-06057A000



# Bench surface protection paper

Bench surface protection paper for all laboratory requirements.

PRODUCT	DIMENSIONS	PACK	CODE	
Plastic-coated paper	48 x 60 cm	100 pcs.	08-CA2000	
Filter paper	50 x 50 cm	500 pcs.	08-656	

# Slide adapter for large samples

For use in conjunction with racks for automatic and manual stainers for staining slides with large samples together with standard slides.

PRODUCT	PACK	CODE	
Slide adapter for large samples	1 pc.	40-400-267	



LIQUID BLOCKE

#### Pap Pen

Deposits a waxy water-repellent film on slides to outline the staining area.

PRODUCT	TIP DIAMETER	PACK	CODE
Liquid Blocker	5 mm	1 pc.	11-100
Liquid Blocker Mini	2 mm	1 pc.	11-100M



# Histology pen

Pen with special permanent ink that remains color-fast during processing to ensure reliable identification of preparations.

Writes on glass, metal, porcelain and plastic.

COLOR	PACK	CODE
Black	12 pcs.	11-50

# **Tube Checker**

Black

Special ink pens for permanent marking of embedding cassettes. The ink is resistant to alcohol and xylene. The pens are equipped with two tips, one fine and one broad.

COLOR	PACK	CODE

1 pc.

11-400

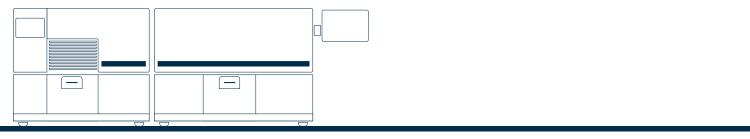


#### Pen with diamond tip

For permanent engraving of slides.

PRODUCT	PACK	CODE
With hexagonal aluminum handle	1 pc.	08-DS2F

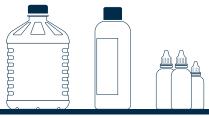




# **Microscope slides**

Cleaned, degreased, high-quality, original Bio-Optica microscope slides; cellophanewrapped and free from dust, dirt and cracks. Resistant to enzyme treatments and microwaves (750-800 Watts). Dimensions: 25.5 x 75.5 mm.





	EDGE	BAND	PACK	CODE
• • • • • • •	Cut Ground 90°- beveled corners 45° Ground 90°- beveled corners 45°	Frosted Neutral Frosted Pink Blue Green White Yellow Orange	2500 pcs. 2500 pcs. 2500 pcs. 2500 pcs. 2500 pcs. 2500 pcs. 2500 pcs. 2500 pcs. 2500 pcs.	09-1000TB 09-1000M 09-1000 09-1010 09-1020 09-1020 09-1030 09-1040 09-1050
•	Ground 90°- beveled corners 45° positively charged	White	72 pcs.	09-3000



# COMPATIBILITY WITH COMMERCIALLY AVAILABLE WRITING SYSTEMS

CODE	HANDWRITING	LASER PRINTER	THERMO PRINTERS	LEICA PRINTER
09-1000MB	V	Х	Х	Х
09-1000	V	V	V	Х
09-1010	V	V	V	Х
09-1020	V	V	V	Х
09-1030	V	V	V	Х
09-1040	V	V	V	Х
09-1050	V	V	V	Х
09-3000	V	V	V	Х

# Microscope slides for big sections

Cleaned, degreased, high-quality, original Bio-Optica microscope slides. Dimensions: 52 mm x 76 mm and thickness: 1-1.1 mm.

EDGE BAND PACK CODE

Ground 90°- beveled corners 45° Double Frosted 1250 pcs. 09-7000







# **SPECIAL STAINS GUIDE**

This section offers a comprehensive range of high-quality staining solutions designed to enhance the visualization of cellular and tissue structures in histology and cytology samples. Our products ensure precise, reliable, and reproducible results, aiding in accurate diagnosis and research. Additionally, you will find detailed information about staining methods, expected results, and product codes to facilitate your selection process.

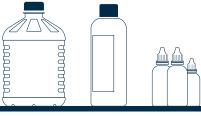
# Manual staining kit

Bio-Optica staining kits have earned ever wider acclaim in Italy and throughout the World for a number of specific reasons, including:

- Quick and easy to use
- Reproducible results
- Predictable cost
- User safety
- Limited environmental impact

Nevertheless, Bio-Optica is committed to continuous improvement of its products and their protocols for use, thanks in part to feedback from our customers, which help us uphold the highest standards of quality for our products.





#### **GENERAL WARNINGS**

#### For best results, please read the following guidelines.

#### Minimum number of tests that can be performed

The number of tests is calculated by assuming reagent consumption of 10 drops per test, which is more than sufficient to cover even medium-large sections. If you wish to use a smaller number of drops when working with small samples, you must reduce the quantity of each reagent in the same proportion in order to avoid imbalances.

#### **Completion time**

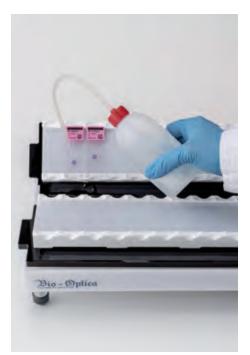
The completion time is calculated according to the duration of the individual steps of which the method consists. It does not include the time taken for de-waxing, hydrating and dehydrating the section.

#### **Essential basic equipment**

To complete the kit, you will need the following essential basic equipment:

- Slide Master, code 15-MEQ0001, for horizontal slide staining
- Spray bottle containing distilled water to perform the steps required by the protocol
  Two series of solvents:
- descending for de-waxing the sections and bringing them to the water and ascending to dehydrate and diaphanize the section before mounting with coverglass.

We recommend the use of BioMount HM (codes 05-BMHM100 or 05-BMHM508) as the mounting medium.





#### **Additional equipment**

The individual instructions indicate which equipment not included in the kit, but usually available in the laboratory, is necessary to complete the kit.

#### Fixatives and embedding media

The protocol times were determined on histological sections of fragments fixed in formalin buffered to pH 7 with phosphate buffer and subsequently embedded in paraffin.

Afog

PRODUCT AND APPLICATION		CODE
Afog Acid Fuchsin Orange G		04-021002
Minimum number of tests that can be performed	100	
Completion time	22 minutes	
Shelf life	2 years	
Storage conditions	15-25°C	
Additional equipment	Not required	

# **Application**

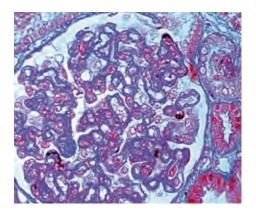
Reference method for highlighting protein deposits in renal biopsy. Recommended fixative: Bouin.

#### Method

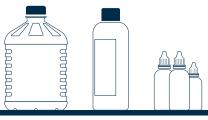
- 1) Bring the section to the distilled water.
- 2) Dispense 5 drops of reagent A and 5 drops of reagent B onto the section: leave to act for 10 minutes.
- 3) Tap water for 5 minutes.
- 4) Dispense 10 drops of reagent C onto the section: leave to act for 5 minutes.
- 5) Wash in distilled water.
- 6) Dispense 10 drops of reagent D onto the section: leave to act for 5 minutes.
- 7) Wash in distilled water.
- 8) Dehydrate rapidly by means of the ascending series of alcohols, stopping for 1 minute in the last absolute; xylene and balsam.

#### KIDNEY

Results



Collagen fibrils	blue
Nuclei	black
Erythrocytes, cytoplasm	pink - orange
Elastic fibers	pale pink - yellow or colorless
Protein deposits	bright red



PRODUCT AND APPLICATION	CODE	AgNOR
AgNOR	04-045801	_
Minimum number of tests that car be performed	12 preparations (up to 4 slides per preparation)	
Completion time	30 minutes	_
Shelf life	1 year	_
Storage conditions	15-25°C	_
Additional equipment	glass rod, jars for washing in distilled water	

#### Application

Method for highlighting argentaffin proteins (100 kD) in the nucleolus organizer region (NOR) on paraffin-embedded sections and smears.

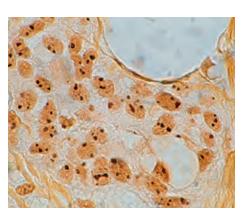
#### Method

- 1) Bring the section to the distilled water.
- 2) Preparation of the work solution: place the slide container in the polystyrene stand. Pour the entire contents of bottle A and the entire contents of a bottle B into the container. Stir briefly with a glass rod previously washed in distilled water.
- 3) Place the section in the solution and incubate in the dark for 30 minutes at room temperature.
- 4) Wash thoroughly in three changes of distilled water.
- 5) Dispense 10 drops of reagent C onto the section: leave to act for 1 minute.
- 6) Wash in distilled water.
- 7) Dehydrate by means of the ascending series of alcohols, xylene and balsam.

# Result AgNOR, argentaffin granules black

#### WARNINGS

- For washing, it is imperative to use top-quality distilled water.
- Do not use Poly-L-Lysine coated slides.
- Do not use metal objects (racks, forceps).
- After mounting, keep the slides in the dark.



#### Results

BREAST

# Alcian Blue pH 2.5

	PRODUCT AND APPLICATION		CODE	
•	Alcian Blue pH 2.5		04-160802	
	Minimum number of tests that can be performed	100		
	Completion time	50 minutes		
	Shelf life	2 years		
	Storage conditions	15-25°C		
	Additional equipment	Not required		

# Application

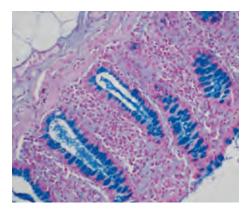
Method indicated for highlighting acid mucopolysaccharides on tissue sections.

#### Method

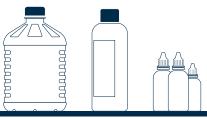
- 1) Bring the section to the distilled water.
- 2) Dispense 10 drops of reagent A onto the section: leave to act for 30 minutes.
- 3) Without washing, drain the slide and dispense 10 drops of reagent B onto the section: leave to act for 10 minutes.
- 4) Wash in distilled water.
- 5) Dispense 10 drops of reagent C onto the slide: leave to act for 5 minutes.
- 6) Wash in distilled water.
- 7) Dehydrate by means of the ascending series of alcohols; xylene and balsam.

#### INTESTINE

Results



Acid mucopolysaccharides	blue - turquoise
Nuclei	red



PRODUCT AND APPLICATION		CODE	Alcian Blue pH 2.5 P.A.S.
Alcian Blue pH 2,5 PAS		04-163802	
Minimum number of tests that can be performed	100		
Completion time	1 hour 25 minutes		
Shelf life	1 year		

#### **Application**

Storage conditions

Additional equipment

Combined method for differentiating acid mucins, neutral mucins and carbohydrates on tissue sections.

2-8°C

Not required

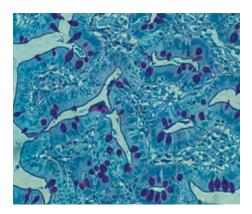
# Method

- 1) Bring the section to the distilled water.
- 2) Dispense 10 drops of reagent A onto the section: leave to act for 30 minutes.
- 3) Without washing, drain the slide and dispense 15 drops of reagent B onto the section: leave to act for 10 minutes.
- 4) Wash for 5 minutes in tap water and for 2 minutes in distilled water.
- 5) Dispense 10 drops of reagent C onto the section: leave to act for 10 minutes.
- 6) Wash in distilled water.
- 7) Dispense 10 drops of reagent D onto the section: leave to act for 20 minutes.8) Wash in distilled water.
- 9) Dispense 10 drops of reagent E onto the section: leave to act for 2 minutes.
- 10) Without washing, drain the slide and dispense 10 drops of reagent F onto the
- section: leave to act for 3 minutes.
- 11) Wash in distilled water.
- 12) Dispense 10 drops of reagent G onto the section: leave to act for 2 minutes.
- 13) Leave to develop in running tap water for 5 minutes.
- 14) Dehydrate in the ascending series of alcohols; xylene and balsam.

#### Result

PAS-positive substances	magenta red
Acid mucopolysaccharides	blue - turquoise
Certain acid mucins and cartilage	from purple to dark blue







# Alcian Yellow - Toluidine blue

•	Alcian Yellow - Toluidine blue for Helicobacter pylori	04-169812
	PRODUCT AND APPLICATION	CODE

Minimum number of tests that can100be performed25 minutesCompletion time25 minutesShelf life2 yearsStorage conditions15-25 °CAdditional equipmentNot required

### Application

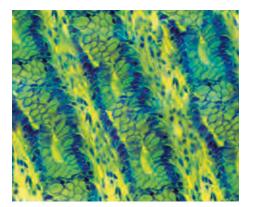
Combined method for highlighting Helicobacter pylori and epithelial mucins on sections of gastric tissue; recommended section thickness 5 microns.

# Method

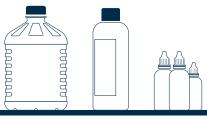
- 1) Bring the section to the distilled water.
- 2) Dispense 10 drops of reagent A onto the section: leave to act for 10 minutes.
- 3) Wash thoroughly in distilled water.
- 4) Dispense 10 drops of reagent B onto the section: leave to act for 5 minutes.
- 5) Wash in tap water for 2 minutes.
- 6) Dispense 10 drops of reagent C onto the slide: leave to act for 5 minutes.
- 7) Wash thoroughly in distilled water.
- 8) Dispense 8 drops of reagent D and 2 drops of reagent E onto the section: leave to act for 3 minutes.
- 9) Wash thoroughly in distilled water.
- 10) Dry in air.
- 11) Dehydrate in alcohol; xylene and balsam.

#### Results

INTESTINE



Helicobacter pylori	blue	
Mucins	yellow	
Background	blue	



PRODUCT AND APPLICATION			CODE	Amylase - Enzymatic digestion
•	Amylase - enzymatic digestion		04-140808	
	Minimum number of tests that can be performed	100		
	Completion time	10 minutes		
	Shelf life	1 year		
	Storage conditions	2-8°C		
	Additional equipment	Vertical jar		

# Application

Removal of glycogen from

- Hepatic tissue, paraffin-embedded sections: digestion on a histological section with a solution of amylase is indicated when you want to remove the glycogen so as to observe only the neutral epithelial mucins. It is the method of choice in liver biopsy.

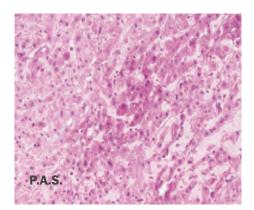
- Muscle tissue: the examination of adjacent cryostat sections, one of which has been treated with amylase, allows qualitative evaluation of the presence of glycogen.

### Method

- 1) Bring the section to the distilled water.
- 2) Bring the amylase solution to room temperature.
- 3) Cover the section with the amylase solution: leave to act for 10 minutes at room temperature.
- 4) Wash the slide several times in distilled water.
- 5) Proceed with the PAS reaction in the normal manner.

LIVER

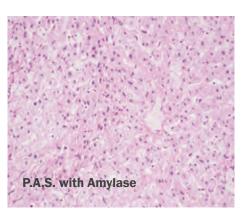
Results



LIVER

# Result

The removal of glycogen can be detected, after PAS reaction, by comparing the sample section with an adjacent section of the same preparation that has not been treated with amylase.



# **Azan Trichrome**

PRODUCT AND APPLICATION

CODE

Azan Trichrome 04-001802

Minimum number of tests that can100be performedCompletion time1 hour 40 minutesShelf life2 yearsStorage conditions15-25 ° CAdditional equipmentVertical histology jar, oven

### Application

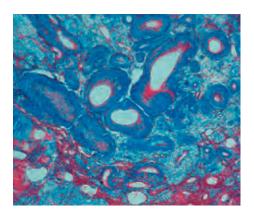
The method of choice for connective tissue, particularly indicated for muscle and glial fiber, collagen, reticulum, glomerular stroma of the kidney, erythrocytes and nuclear chromatin on histological sections.

#### Method

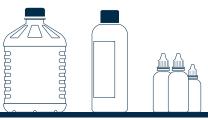
- 1) Bring the section to the distilled water.
- 2) Incubate the section in reagent A in an oven at 56°C for 30 minutes, then remove from the oven and wait for 5 minutes. Retrieve the stain and transfer it to bottle A without filtering it.
- 3) Wash in distilled water.
- 4) Dispense 10 drops of reagent B onto the section: leave to act for 1 minute.
- 5) Drain on filter paper, then dispense 10 drops of reagent C onto the section: leave to act for 1 minute.
- 6) Drain on filter paper, then dispense 10 drops of reagent D onto the section: leave to act for 30 minutes.
- 7) Drain on filter paper and without washing, dispense 10 drops of reagent E onto the section: leave to act for 30 minutes.
- 8) Wash quickly in 95° ethanol. Dehydrate in the ascending series of alcohols; xylene and balsam.

Results

OVARY



ncoun	
Collagen, reticulum, basophilic cytoplasmic granules of the pituitary gland, juxtaglomerular granules and glomerular stroma of	blue
the kidney	
Neurofibrils (glia)	reddish
Muscle	orange
Nuclei, erythrocytes and acidophilic	red
granules of the pituitary gland	
Cytoplasmic granules of the delta	blue
cells of the pituitary gland	



**Bielschowsky** 

# **Staining and mounting**

PRODUCT AND APPLICATION		CODE
Bielschowsky for neurofibrils		04-040805
Minimum number of tests that can be performed	100	
Completion time	45 minutes	
Shelf life	1 year	
Storage conditions	2-8°C	

oven, 50 ml Coplin jar, glass rod

#### Application

Additional equipment

The method of choice for viewing neurofibrils, axons, dendrites and senile plaques. Usable on sections fixed in 10% formalin and embedded in paraffin, having a thickness of 8 – 10  $\mu m.$ 

#### Method

- 1) Bring the section to the distilled water.
- 2) Place the slide in a humid chamber, dispense 10 drops of reagent A onto the section; close the lid and incubate in the oven at 40°C for 15 minutes.
- 3) Remove the slide from the humid chamber and wash the section thoroughly in distilled water.
- 4) Return the slide to the humid chamber and dispense 10 drops of reagent B onto the section: close the lid and incubate in the oven at 50/55 °C for 20 minutes. During this incubation period, prepare the reducing solution as follows: dispense 50 ml of distilled water into a Coplin jar and add 20 drops of reagent C, 8 drops of reagent D, 8 drops of reagent E and 8 drops of reagent F. Stir briefly with a glass rod.
- 5) Without washing, drain the slide and place it in the reducing solution: leave to act
- for 1-2 minutes.
- 6) Wash twice in distilled water.
- 7) Dispense 10 drops of reagent G onto the section: leave to act for 3 minutes.
- 8) Wash twice in distilled water.
- 9) Dehydrate by means of the ascending series of alcohols; xylene and balsam.

#### Result

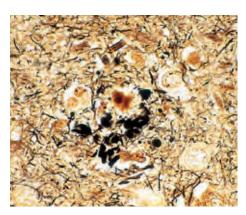
Neurofibrils and senile plaques	black
Background	from yellow to brown

#### WARNINGS

The success of the reaction depends on rigorous adherence to the following rules:

- Always use good-quality, totally chlorine-free distilled or deionized water.
- Use only rigorously clean glassware or plastic ware.
- Never bring metal objects (forceps etc.) into contact with the solutions.

#### CEREBRAL CORTEX



# **Brown - Brenn**

PRODUCT AND APPLICATION

Brown - Brenn for bacteria

Minimum number of tests that can	100
be performed	
Completion time	8 minutes
Shelf life	2 years
Storage conditions	15-25°C
Additional equipment	Not required

CODE

04-100807

#### **Application**

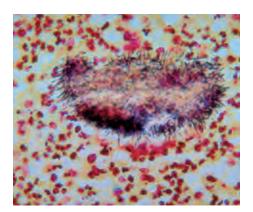
Method for differentiating Gram-positive and Gram-negative bacteria on histological sections and smears.

#### Method

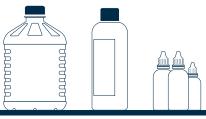
- 1) Bring the section to the distilled water.
- 2) Dispense 8 drops of reagent A and 2 drops of reagent B onto the section: leave to act for 1 minute.
- 3) Wash in distilled water.
- 4) Dispense 10 drops of reagent C onto the section: leave to act for 3 minutes
- 5) Wash in distilled water and dry the slide with filter paper.
- 6) Dispense 10 drops of reagent D onto the section: leave to act for 1 minute.
- 7) Drain without washing and dispense 10 drops of reagent E onto the section: leave to act for 1 minute
- 8) Wash in distilled water and dry the slide with filter paper.
- 9) Dispense 10 drops of reagent F onto the section: leave to act for 1 minute.
- 10) Drain without washing and dispense 10 drops of reagent G onto the section: leave to act for 30 seconds.
- 11) Xylene and balsam.

#### Results

OVARY



Gram-positive bacteria	blue
Gram-negative bacteria	red
Actinomycetes (Nocardia)	blue
Nuclei	red
Other tissue elements	yellow



	PRODUCT AND APPLICATION		CODE	<b>Diastase - Enzymatic digestion</b>
•	Diastase for enzymatic digestion		04-140805	-
	Minimum number of tests that can	40		
	be performed			
	Completion time	30 minutes		-
	Shelf life	1 year		-
	Storage conditions	2-8°C		-
	Additional equipment	vertical jar		-

### Application

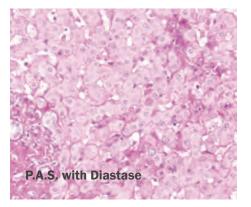
Digestion on a histological section with a solution of diastase is always indicated when you want to remove the glycogen so as to observe only the neutral epithelial mucins. It is the method of choice in liver biopsy.

# Method

- 1) Bring the section to the distilled water.
- 2) Bring the diastase solution to room temperature.
- 3) Incubate the slide at room temperature for 30 minutes.
- 4) Wash the slide several times in distilled water.
- 5) Proceed with the PAS reaction in the normal manner.

Results

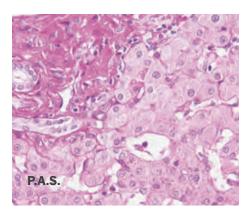
LIVER



LIVER

#### Result

The removal of glycogen can be detected, after PAS reaction, by comparing the sample section with an adjacent section of the same preparation that has not been treated with diastase.



# **Colloidal iron**

PRODUCT AND APPLICATION		CODE 04-180809	
Colloidal iron, method for acid mud	cins		
Minimum number of tests that can be performed	100		
Completion time	1 hour 35 minutes		
Shelf life	2 years		
Storage conditions	15-25°C		
Additional equipment	50 ml vertical histology jar, gradua	ted cylinder and glass	

rod

#### Application

Indicated method for viewing acid mucins.

Specificity: the reaction shows acid mucins (sialomucins and sulfomucins) whose acid groups, at the reaction pH, take anionic form and are therefore capable of forming a stable complex with positive trivalent iron.

# Method

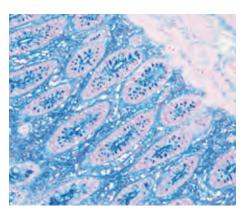
- 1) Bring the section to the distilled water.
- 2) Dispense 10 drops of reagent A onto the section: leave to act for 2 minutes.
- 3) Prepare the humid chamber as follows: soak the disk of filter paper with approximately 1 ml of distilled water, insert the slide and dispense 5 drops of reagent B and 5 drops of reagent C onto the section, then close the lid and incubate for 1 hour.
- 4) Without washing, drain the slide and dispense 10 drops of reagent D onto it: leave to act for 1 minute. Drain and repeat.
- 5) Without washing, drain the slide and dispense 10 drops of reagent E onto it: leave to act for 1 minute. Drain and repeat.
- 6) Drain the slide.
- 7) Prepare the potassium ferrocyanide solution as follows: pour the entire contents of a bottle F into a 50 ml Coplin jar. Add, in order, 30 ml of distilled water and 4 ml of reagent G. Stir briefly. Immerse the section for 10 minutes.
- 8) Wash thoroughly in distilled water.
- 9) Dispense 10 drops of reagent H onto the section: leave to act for 5 minutes.
- 10) Wash in distilled water.
- 11) Dehydrate in 95° and absolute ethanol; xylene and balsam.

#### Result

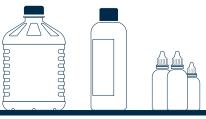
Results

Acid mucins	blue
Cellular nuclei	red

INTESTINE



p\_68/139



PRODUCT AND APPLICATION		CODE	Fouchet-Van Gieson
Fouchet-Van Gieson for bilirubin		04-121872	-
Minimum number of tests that can be performed	100		
Completion time	35 minutes		_
Shelf life	2 years		_
Storage conditions	15-25°C		_

#### **Application**

For highlighting bilirubin pigment on tissue sections.

# Method

- 1) Bring the sections to the distilled water.
- 2) Dispense 5 drops of reagent A onto the section and add 5 drops of reagent B, leave

Not required

to act for 5 minutes.

Additional equipment

- 3) Wash in distilled water.
- 4) Dispense 10 drops of reagent C onto the section: leave to act for 7 minutes.
- 5) Without washing, drain the slide and dry it first in filter paper, then in the air for 5 minutes.
- 6) Absolute alcohol for 15 seconds, xylene and balsam.

#### **BILIRUBIN DEPOSITS**





# **Paraldehyde Fuchsin**

PRODUCT AND APPLICATION

Paraldehyde Fuchsin - Gomori 04-045872

CODE

Minimum number of tests that can100be performedCompletion time1 hour 15 minutesShelf life2 yearsStorage conditions15-25 °CAdditional equipmentNot required

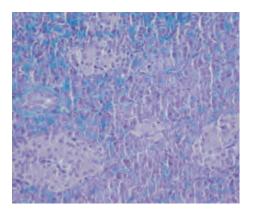
#### Application

For viewing elastic fibers and secretory granules in alpha and beta cells of the islets of Langerhans of the endocrine pancreas.

### Method

- 1) Bring the sections to the distilled water.
- 2) Dispense 5 drops of reagent A onto the section and add 5 drops of reagent B, leave to act for 10 minutes.
- 3) Wash in distilled water.
- 4) Dispense 10 drops of reagent C onto the section, leave to act for 5 minutes.
- 5) Wash in distilled water.
- 6) Dispense 10 drops of reagent D onto the section, leave to act for 5 minutes.
- 7) Without washing, drain the slide and place it in the humid chamber, then dispense 10 drops of reagent E onto the section and leave to act for 20 minutes.
- 8) Drain the slide and dispense 10 drops of reagent F onto the section, then leave to act for 10 minutes.
- 9) Wash the slide in distilled water.
- 10) Dispense 10 drops of reagent G onto the section, leave to act for 10 minutes.
- 11) Wash in distilled water.
- 12) Dispense 10 drops of reagent H onto the section, leave to act for 30 seconds.
- 13) Wash in distilled water, dehydrate in 95 and absolute alcohol, xylene and balsam.

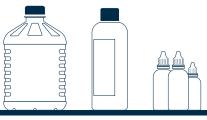
#### Results



#### Results

PANCREAS

Pancreatic beta-cell granules	dark violet
Cellular nuclei	dark violet
Connective tissue	red
Tessuto connettivo	green



PRODUCT AND APPLICATION		CODE	
Giemsa for Helicobacter pylori		04-090803	
Minimum number of tests that can be performed	75		
Completion time	1 hour		
Shelf life	2 years		
Storage conditions	15-25°C		
Additional equipment	Graduated cylinder		

#### **Application**

Method for viewing Helicobacter Pylori on sections from gastric biopsy. The qualitative and quantitative composition of the stain and the accurate differentiation make it possible to identify bacteria selectively on a particularly clean background.

### Method

Result

Nuclei

Cytoplasm

Helicobacter pylori

- 1) De-wax the sections and bring them to the water.
- 2) Prepare the buffer solution: take 5 ml of solution from bottle B and dilute in a ratio of 1:10.
- Use the solution thus obtained to prepare the working Giemsa solution.
- 3) Prepare the working Giemsa solution: take 10 ml of reagent A and top up to 40 ml with the previously prepared buffer solution.

blue, in the characteristic gullwing shape

- 4) Place the solution in the jar and immerse the sections in it for 30 minutes.
- 5) Drain and, without washing, place the section in reagent C for 15 seconds.

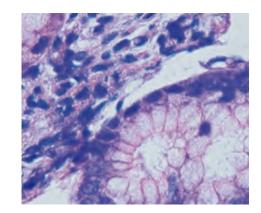
blue

pink

- 6) Repeat step 5 with reagents D and E.
- 7) Diaphanize in xylene and mount with balsam.

#### GASTRIC MUCOSA

Giemsa



# **Gordon-Sweet**

PRODUCT AND APPLICATION

Gordon-Sweet - for reticulum

Minimum number of tests that can	100
be performed	
Completion time	40 minutes
Shelf life	1 year
Storage conditions	2-8°C
Additional equipment	Not required

CODE

04-040802

# Application

The method of choice for viewing argyrophilic reticular fibers of connective tissue.

#### Method

- 1) Bring the section to the distilled water.
- 2) Dispense 5 drops of reagent A and 5 drops of reagent B onto the section: leave to act for 5 minutes.
- 3) Wash in distilled water.
- 4) Dispense 10 drops of reagent C onto the section: leave to act for 1 minutes.
- 5) Wash twice in distilled water.
- 6) Dispense 10 drops of reagent D onto the section: leave to act for 3 minutes.
- 7) Wash twice in distilled water.
- 8) Dispense 10 drops of reagent E onto the section: leave to act for 3 minutes.
- 9) Wash in distilled water.
- 10) Dispense 10 drops of reagent F onto the section: leave to act for 5 minutes.
- 11) Wash twice in distilled water.
- 12) Dispense 10 drops of reagent G onto the section: leave to act for 2 minutes.
- 13) Wash in distilled water.
- 14) Dispense 10 drops of reagent H onto the section: leave to act for 2 minutes.
- 15) Wash in distilled water.
- 16) Dispense 10 drops of reagent I onto the section: leave to act for 5 minutes.
- 17) Dehydrate by means of the ascending series of alcohols; xylene and balsam.

#### Result

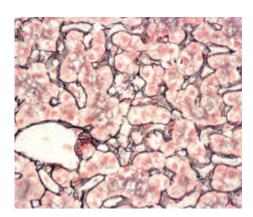
LIVER

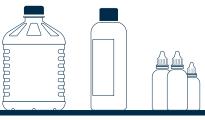
Reticular and nerve fibers	black
Nuclei	red, pink

#### WARNINGS

The success of the reaction depends on rigorous adherence to the following rules:

- Always use good-quality, totally chlorine-free distilled or deionized water.
- Use only rigorously clean glassware.
- Avoid depositing dust on the sections.
- Never bring metal objects (forceps etc.) into contact with the solutions.





Gram

# **Staining and mounting**

PRODUCT AND APPLICATION	CODE	
Gram for bacteria	04-100802	2
Minimum number of tests that can	100	
be performed		
Completion time	40 minutes	
Shelf life	2 years	
Storage conditions	15-25°C	
Additional equipment	3 vertical glass histology jars, funnel, filter, ove	n

#### **Application**

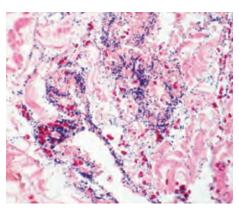
Method for differentiating Gram-positive and Gram-negative bacteria on histological sections, smears and tissue apposition.

### Method

- 1) Bring the section to the distilled water.
- 2) Pour the contents of bottle A into a vertical histology jar, place the slide in it and incubate at 56-58°C for 15 minutes; retrieve the solution and transfer it to bottle A, filtering it through filter paper.
- 3) Wash in distilled water.
- 4) Dispense 10 drops of reagent B onto the section: leave to act for 3 minutes.
- 5) Drain the slide and, without washing it, dispense 10 drops of solution C onto it: leave to act for 3 minutes.
- 6) Wash in distilled water and dry the slide first in filter paper, then in the air for 10 minutes.
- 7) Pour the contents of bottle D into a vertical histology jar: stir the slide in it for 1 minute; retrieve the solution and transfer it to bottle D, filtering it through filter paper.
- 8) Repeat step 7 with reagent E.
- 9) Xylene and balsam.

# NECROTIZING FASCIITIS







#### Gram for microbiology

 PRODUCT AND APPLICATION
 CODE

 Gram for microbiology
 04-100803

 with basic fuchsin as counterstain solution
 04-100804

 Completion time
 6 minutes

 Shelf life
 2 years

 Storage conditions
 15-25 ° C

 Additional equipment
 graduated cylinder, glass rod, oven

#### **Application**

Method for differentiation of gram-positive and gram-negative bacteria in fixed smears. This method is often used to assess suitability of specimen for culture.

#### Method

- 1) Fix air-dried smears using one of the following techniques:

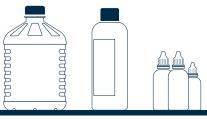
   a Heat fix by passing the slide through a low flame 2-3 times. Cool the slide at room temperature before staining
   b Methanol fix, immerse the slide in absolute methanol for 1 2 minutes and rinse with distilled water before staining
- 2) Cover specimens with reagent A (Crystal-violet Hucker solution), leave to act 1 minute.
- 3) Drain the slide and flood briefly with reagent B (Lugol solution).
- 4) Cover completely with reagent B (Lugol solution) and leave to act 1 minute.
- 5) Wash with distilled water.
- 6) Cover completely with reagent C (Decolorizing solution): leave to act 1 minute.
- 7) Wash with distilled water.
- 8) Cover completely with reagent D (Safranin solution): Leave to act 1 minute.
- 9) Wash carefully with distilled water.
- 10) Air dry.

#### Results

GRAM-NEGATIVE BACTERIA



Gram-positive bacteria	Blue-violet	
Gram-negative bacteria	Pink-Red	



Grocott

#### **Staining and mounting**

PRODUCT AND APPLICATION		CODE
Grocott for fungi		04-043823
Minimum number of tests that can be performed	120	
Completion time	1 hour 50 minutes	
Shelf life	1 year	
Storage conditions	2-8°C	
Additional equipment	graduated cylinder, glass rod, oven	

#### Application

Method used for viewing fungi on a tissue section.

#### Method

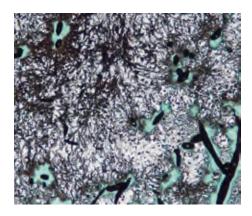
- 1) Bring the section to the distilled water.
- 2) Dispense 10 drops of reagent A onto the section, leave to act for 20 minutes. Wash in running water for a few seconds.
- 3) Dispense 10 drops of reagent B onto the section: leave to act for 1 minute. Wash in tap water for 5 minutes.
- 4) Wash in four changes of distilled water.
- 5) Pour 17 ml of distilled water into a slide container and add: 20 drops of reagent C, 10 drops of reagent D, 20 drops of reagent E. Stir briefly with a glass rod washed in distilled water.
- 6) Place the slide in the container and incubate for 1 hour in an oven at 60°C.
- 7) Remove the container from the oven and leave to cool for 10 minutes. Wash in 6 changes of distilled water.
- 8) Dispense 10 drops of reagent F onto the section; leave to act for 3 minutes. Rinse in distilled water.
- 9) Dispense 10 drops of reagent G onto the section; leave to act for 5 minutes. Wash in tap water.
- 10) Dispense 10 drops of reagent H onto the section; leave to act for 30 seconds.
- 11) Dehydrate by means of the ascending series of alcohols; xylene and balsam.

Result			
Fungi	clearly outlined in black		
Mucins	dark gray		
Background	green		

#### WARNINGS

The success of the reaction depends on rigorous adherence to the following rules:

- Avoid contaminating the section and microscope slide with non-pathogenic fungi (handle only with gloves, do not leave the preparation exposed to air).
- Always use recently distilled water.
- Use only rigorously clean glassware.
- Avoid depositing dust on the sections.
- Never bring metal objects (forceps etc.) into contact with the solutions.



LUNG



#### **Grocott for microwave oven**

PRODUCT AND APPLICATION

Grocott MW for fungi 04-043823W

CODE

 Minimum number of tests that can
 120

 be performed
 50 minutes

 Completion time
 50 minutes

 Shelf life
 1 year

 Storage conditions
 2-8 ° C

 Additional equipment
 graduated cylinder, glass rod, oven

#### Application

Method used for viewing fungi on a tissue section.

#### Method

- 1) Bring the section to the distilled water.
- 2) Dispense 10 drops of reagent A onto the section, leave to act for 20 minutes. Wash in running water for a few seconds.
- 3) Dispense 10 drops of reagent B onto the section: leave to act for 1 minute. Wash in tap water for 5 minutes.
- 4) Wash in four changes of distilled water.
- 5) Pour 40 ml of distilled water into a 50 ml Coplin jar and add: 30 drops of reagent C, 15 drops of reagent D, 20 drops of reagent E. Stir briefly with a glass rod washed in distilled water.
- 6) Put the slides in the jar and place in a microwave oven at 500W for 1 minute.
- 7) Remove the jar from the oven and leave to cool for 5 minutes. Wash in 6 changes of distilled water.
- 8) Dispense 10 drops of reagent F onto the section; leave to act for 3 minutes. Rinse in distilled water.
- 9) Dispense 10 drops of reagent G onto the section; leave to act for 5 minutes. Wash in tap water.
- 10) Dispense 10 drops of reagent H onto the section; leave to act for 30 seconds.
- 11) Dehydrate by means of the ascending series of alcohols; xylene and balsam.
- II) Denyulate by means of the ascending series of alcohols, sylene and balsam

Result	Result			
Fungi	clearly outlined in black			
Mucins	dark gray			
Background	green			

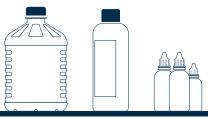
#### WARNINGS

The success of the reaction depends on rigorous adherence to the following rules:

- Avoid contaminating the section and microscope slide with non-pathogenic fungi (handle only with gloves, do not leave the preparation exposed to air).
- Always use recently distilled water.
- Use only rigorously clean glassware.
- Avoid depositing dust on the sections.
- Never bring metal objects (forceps etc.) into contact with the solutions.

#### Results

NASAL POLYP



	PRODUCT AND APPLICATION		CODE	Silver impregnation
•	Silver impregnation staining for re	ticulum	04-040801	-
	Minimum number of tests that can be performed	100		
	Completion time	35 minutes		-
	Shelf life	1 year		_
	Storage conditions	2-8°C		-
	Additional equipment	Not required		

#### Application

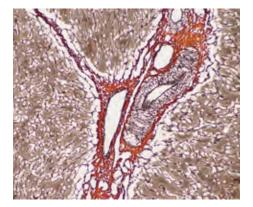
The method of choice for viewing argyrophilic reticular fibers of connective tissue.

#### Method

- 1) Bring the section to the distilled water.
- 2) Dispense 5 drops of reagent A and 5 drops of reagent B onto the section: leave to act for 5 minutes.
- 3) Wash in distilled water.
- 4) Dispense 10 drops of reagent C onto the section: leave to act for 3 minutes.
- 5) Wash twice in distilled water.
- 6) Dispense 10 drops of reagent D onto the section: leave to act for 3 minutes.
- 7) Wash twice in distilled water.
- 8) Dispense 10 drops of reagent E onto the section: leave to act for 3 minutes.
- 9) Wash in distilled water.
- 10) Dispense 10 drops of reagent F onto the section: leave to act for 5 minutes.
- 11) Wash twice in distilled water.
- 12) Dispense 10 drops of reagent G onto the section: leave to act for 5 minutes.
- 13) Wash in tap water for 5 minutes.
- 14) Dehydrate by means of the ascending series of alcohols; xylene and balsam.

#### Results

LIVER



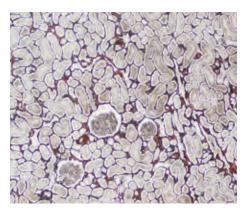
KIDNEY

Result	
Reticular and nerve fibers	black
Connective tissue	brown
Collagen	yellow

#### WARNINGS

The success of the reaction depends on rigorous adherence to the following rules:

- Always use good-quality, totally chlorine-free distilled or deionized water.
- Use only rigorously clean glassware.
- Avoid depositing dust on the sections.
- Never bring metal objects (forceps etc.) into contact with the solutions.



#### Luxol fast blue

 PRODUCT AND APPLICATION
 CODE

 Luxol fast blue, Klüver-Barrera method
 04-200812

 Minimum number of tests that can
 100

 be performed
 20 minutes + overnight

 Completion time
 20 minutes + overnight

 Shelf life
 2 years

 Storage conditions
 15-25 °C

Not required

#### **Application**

Additional equipment

Method indicated for showing myelin and phospholipids on histological sections.

#### Method

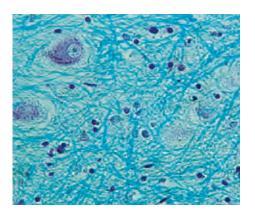
- 1) De-wax the section and bring it to the 95° ethanol.
- 2) Prepare the humid chamber by wetting the filter in the Petri dish with distilled water, place the slide in the rack and then dispense 10 drops of reagent A onto the section; close the lid of the dish immediately and incubate in an oven at 56°C overnight.
- 3) Remove the slide from the humid chamber and wash in 95° ethanol (the crystallized residues of reagent A must also dissolve).
- 4) Wash in distilled water.
- 5) Dispense 10 drops of reagent B onto the section: leave to act for 30 seconds.
- 6) Differentiate in 70° ethanol until the myelinated fibers appear in blue against an almost colorless background (if differentiation proves difficult, repeat step 5 for 30 seconds and put the preparation in 70° ethanol again).
- 7) Wash thoroughly in distilled water (at least 2 changes).
- 8) Prepare the humid chamber; dispense 10 drops of reagent C and 5 drops of reagent D onto the preparation, then incubate at 56°C for 20 minutes.
- 9) Differentiate the preparation in 95° ethanol until the Nissl substance turns pale pink.
- 10) Dehydrate in absolute ethanol; xylene and balsam.

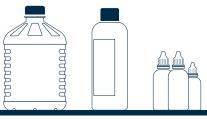
#### Result

Results

Myelin	turquoise blue
Neurons and glial nuclei	from pink to violet
Nissl substance	pale pink

#### CEREBELLUM





PRODUCT AND APPLICATION		CODE	Mallory's Trichrome
Mallory's Trichrome		04-020802	
Minimum number of tests that can be performed	100		
Completion time	20 minutes		
Shelf life	2 years		
Storage conditions	15-25°C		
Additional equipment	Not required		

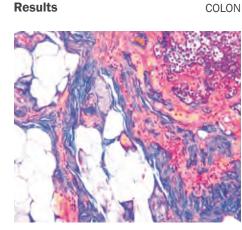
#### **Application**

The standard method for viewing connective tissue on histological sections; particularly indicated for highlighting collagen, reticulum, cartilage, bone and amyloid.

#### Method

- 1) Bring the section to the distilled water.
- 2) Dispense 10 drops of reagent A onto the section: leave to act for 10 minutes.
- 3) Wash in distilled water.
- 4) Dispense 10 drops of reagent B onto the section: leave to act for 2 minutes.
- 5) Wash quickly in tap water (2-3 seconds) and dispense 10 drops of reagent C onto the section: leave to act for 5 minutes.
- 6) Without washing, drain the slide and dispense 10 drops of solution D onto the section: leave to act for 1 minute.
- 7) Wash in distilled water and dehydrate rapidly by means of the ascending series of alcohols, stopping for 1 minute in the last absolute, xylene and balsam.

Result	
Nuclei, neurofibrils, myoglia,	red
cartilage and bone tissue	
Collagen fibrils	blue
Myelin	golden yellow
Elastic fibers	pale pink - yellow or colorless
Erythrocytes	yellow



#### **Masson Fontana**

PRODUCT AND APPLICATION

Masson Fontana for melanin

Minimum number of tests that can	100
be performed	
Completion time	45 minutes + overnight
Shelf life	1 year
Storage conditions	2-8°C
Additional equipment	Not required

CODE

04-041822

#### Application

The method of choice for viewing melanin pigment on sections of histological tissue.

#### Method

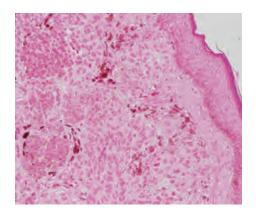
- 1) Bring two slides of the same preparation to distilled water.
- 2) Use one of the two slides as a control. Perform steps 3-4 on the control section only.
- 3) Dispense 10 drops of reagent B and 10 drops of reagent C onto the control slide: leave to act for 20 minutes and then wash in distilled water.
- 4) Dispense 10 drops of reagent D onto the control slide: leave to act for 5 minutes and then wash in distilled water.
- 5) Prepare the humid chamber and place the two slides (sample and control) in it, then dispense 10 drops of reagent A onto each section, close the lid of the humid chamber and leave overnight.
- 6) Wash the incubated sections in distilled water and dispense 10 drops of reagent E onto them: leave to act for 5 minutes.
- 7) Wash in distilled water.
- 8) Dispense 10 drops of reagent F onto the control slide and the sample slide: leave to act for 10 minutes.
- 9) Wash in distilled water.
- 10) Dehydrate by means of the ascending series of alcohols, xylene and balsam.

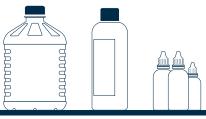
#### Result

Results

Result	
Melanin pigment	brick red - black in the section under examination; absent in the
	control section (the presence of black precipitate on the control
	section indicates a false positive)
Nuclei	pink

SKIN





	PRODUCT AND APPLICATION	D APPLICATION CODE	Ē	Masson Trichrome	
•	Masson Trichrome with aniline blu	ie	04-0	10802	_
	Minimum number of tests that can be performed	100			
	Completion time	35 minutes			
	Shelf life	2 years			
	Storage conditions	15-25°C			

#### Application

Additional equipment

The method of choice for connective tissue, particularly indicated for gametes, nuclei, neurofibrils, glia, collagen, keratin, intracellular fibrils and negative images of the Golgi apparatus.

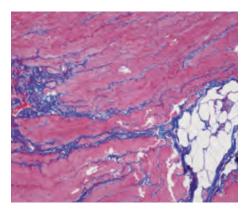
Not required

#### Method

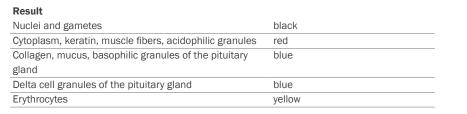
- 1) Bring the section to the distilled water.
- 2) Dispense 6 drops of reagent A onto the section and add 6 drops of reagent B: leave to act for 10 minutes.
- 3) Without washing, drain the slide and dispense 10 drops of solution C onto the section: leave to act for 4 minutes.
- 4) Wash quickly (3-4 seconds) in distilled water, leaving the section yellow in color, and dispense 10 drops of solution D onto the slide: leave to act for 4 minutes.
- 5) Wash in distilled water and dispense 10 drops of solution E onto the section: leave to act for 10 minutes.
- 6) Without washing, drain the slide and dispense 10 drops of solution F onto it: leave to act for 5 minutes.
- 7) Wash in distilled water and dehydrate rapidly by means of the ascending series of alcohols, pausing for 1 minute in the last absolute: xylene and balsam.

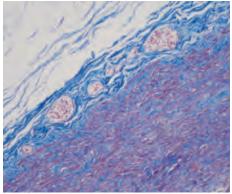
#### Results

#### STOMACH



ARTERY







#### **Masson - Goldner Trichrome**

PRODUCT AND APPLICATION		CODE
Masson-Goldner Trichrome with lig	sht green	04-011802
Minimum number of tests that can be performed	100	
Completion time	35 minutes	
Shelf life	2 years	

15-25°C

Not required

#### **Application**

Storage conditions

Additional equipment

The method of choice for connective tissue, indicated for highlighting gametes, nuclei, neurofibrils, glia, collagen, keratin, intracellular fibrils and negative images of the Golgi apparatus.

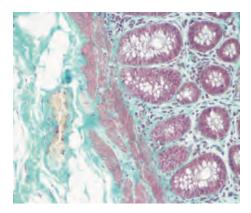
Particularly indicated for black and white micro-photography.

#### Method

Results

- 1) Bring the section to the distilled water.
- 2) Dispense 6 drops of reagent A onto the section and add 6 drops of reagent B : leave to act for 10 minutes.
- 3) Without washing, drain the slide and dispense 10 drops of solution C onto the section: leave to act for 4 minutes.
- 4) Wash quickly (3-4 seconds) in distilled water and dispense 10 drops of solution D onto the slide: leave to act for 4 minutes.
- 5) Wash in distilled water and dispense 10 drops of solution E onto the section: leave to act for 10 minutes.
- 6) Without washing, drain the slide and dispense 10 drops of solution F onto it: leave to act for 5 minutes.
- 7) Wash in distilled water and dehydrate rapidly by means of the ascending series of alcohols, leaving for 1 minute in the last absolute: xylene and balsam.

#### COLON

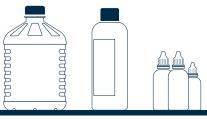


Result

Nuclei and gametes	black
Cytoplasm, keratin, muscle fibers, acidophilic granules	red
Collagen, mucus, basophilic granules of the pituitary gland	green
Delta cell granules of the pituitary gland	green
Erythrocytes	yellow

#### COLON

#### p\_**82**/139



	PRODUCT AND APPLICATION		CODE	May Grünwald Giemsa
•	May Grünwald Giemsa for sections	;	04-081802	
	Minimum number of tests that can be performed	100		
	Completion time	35 minutes		-
	Shelf life	2 years		-
	Storage conditions	15-25°C		-

#### Application

Additional equipment

The method of choice for differentiating cell types and highlighting parasites on tissue sections; particularly indicated for lymphohematopoietic tissue. This stain is often used for identifying endothelial reticulum.

Graduated cylinder

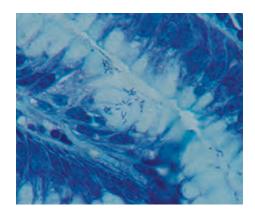
#### Method

- 1) De-wax the section and bring it to the 70° ethanol.
- 2) Prepare the buffer solution: Pour 20 ml of distilled water into the attached container and add 10 drops of concentrated solution B. The diluted solution thus obtained will be designated "buffer solution B".
- 3) Dispense 10 drops of buffer solution B onto the section: leave to act for 2 minutes.
- 4) Drain the slide and dispense 10 drops of reagent A and 5 drops of buffer solution B onto it: leave to act for 5 minutes.
- 5) Pipette 10 ml of buffer solution B and wash the slide thoroughly with it.
- 6) Dispense 10 drops of reagent C and 10 drops of buffer solution B into the dish, stir, place on the slide and leave to act for 12 minutes.
- 7) Differentiate in: 95° ethanol for 10 seconds, absolute ethanol for 30 seconds; absolute ethanol for 30 seconds.
- 8) Xylene and balsam.

# Result Nuclei blue Basophilic cytoplasm from sky blue to dark blue Acidophilic cytoplast pink Bacteria blue

#### Results

#### STOMACH



#### **Mucicarmine**

#### Application

Method indicated for highlighting acid mucopolysaccharides of epithelial nature (mucins) on histological sections.

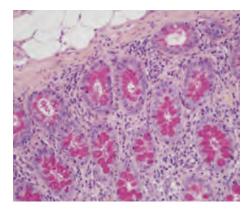
The use of Mucicarmine is of relative specificity, in fact mucins deriving from fibroblasts are generally weakly highlighted.

#### Method

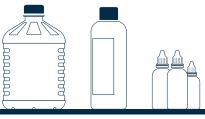
- 1) Bring the section to the distilled water.
- 2) Dispense 10 drops of reagent A onto the section; leave to act for 5 minutes.
- 3) Wash in distilled water.
- 4) Leave to develop in running water for 5 minutes.
- 5) Pipette 0.5 ml of distilled water into the dish, add 10 drops of reagent
- B, stir and transfer the mixture thus obtained to the slide: leave to act for 30 minutes.
- 6) Wash in distilled water.
- 7) Dispense 10 drops of reagent C onto the section: leave to act for 1 minute.
- 8) Wash in distilled water.
- 9) Dehydrate rapidly by means of the ascending series of alcohols, stopping in the last absolute; xylene and balsam.

#### COLON

**Results** 



Mucins	from dark pink to red	
Nuclei	blue – violet	
Other components	orange	



	PRODUCT AND APPLICATION		CODE	Nitro blue tetrazolium
•	Nitroblue tetrazolium		04-253031	
	Minimum number of tests that can be performed	15		
	Completion time	30 minutes		-
	Shelf life	2 years		-
	Storage conditions	2-8°C		-
	Additional equipment	Oven		-

#### Application

Post mortem, infarcted areas of the myocardium undergo a series of changes that are visible in sequence. In the first 6-12 hours after the acute episode, the myocardial infarction is generally neither macroscopically nor microscopically detectable. The ischemic muscle can, however, be highlighted, showing the loss of its oxidative activity with nitroblue tetrazolium staining on fresh sample: the infarcted area remains unstained.

#### Method

- 1) To obtain 150 ml of ready-to-use solution: pour the entire contents of reagents A, B and C into a container of appropriate size and capacity. Stir briefly.
- 2) Immerse the sample of heart in the solution obtained and incubate at 37 °C for 20-30 minutes.
- 3) Wash in tap water and observe the sample: the infarcted area appears pale, not stained.



Oil red O

PRODUCT AND APPLICATION

CODE

04-220923

Minimum number of tests that can	100	

be performed	
Completion time	25 minutes
Shelf life	2 years
Storage conditions	15-25°C
Additional equipment	Glass histology jar with lid

#### Application

Oil red O

Method indicated for highlighting lipids on cryostat sections of tissue having a thickness of 5  $\mu\text{m}.$ 

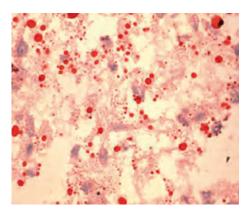
Fixation: you are advised to use saline formalin or Baker fixative in order to make the phospholipids less soluble.

#### Method

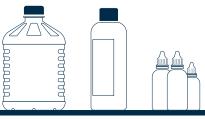
- 1) Bring the section to the distilled water.
- 2) Place the reagent A in the jar and immerse the section in it for 20 minutes.
- 3) Wash briefly in tap water.
- 4) Drain and dispense 10 drops of reagent B onto the section: leave to act for 30 seconds.
- 5) Leave to develop in tap water for 3 minutes.
- 6) Drain and mount with aqueous mounting medium.

#### ADIPOSE TISSUE

**Results** 



Fatty acids bright	red
Nuclei	blue



Orcein

#### **Staining and mounting**

PRODUCT AND APPLICATION	CODE
Orcein for elastic fibers	04-055802
Minimum number of tests that can be performed	100
Completion time	30 minutes
Shelf life	2 years
Storage conditions	15-25°C
Additional equipment	Not required

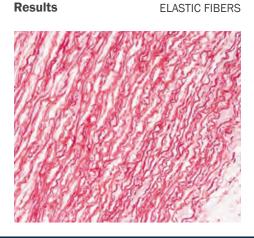
#### Application

Identification of elastic fibers on tissue sections.

#### Method

- 1) Bring the section to the distilled water.
- 2) Dispense 5 drops of reagent A and 5 drops of reagent B onto the section. Leave to act for 4 minutes.
- 3) Wash in distilled water.
- 4) Dispense 10 drops of reagent C onto the section. Leave to act for 1 minute.
- 5) Wash in distilled water.
- 6) Prepare the humid chamber as follows: soak the disc of filter paper with 20 drops of reagent A, insert the slide in the humid chamber and dispense 10 drops of reagent E onto the section. Close the lid and incubate for 20 minutes.
- 7) Wash in distilled water.
- 8) Dispense 10 drops of reagent F onto the section. Leave to act for 2 minutes.
- 9) Wash in running water for 1 minute.
- 10) Dehydrate by means of the ascending series of alcohols, xylene and balsam.

Result	
Elastic fibers	from dark brown to dark purple
Background	almost colorless





#### P.A.S. Periodic Acid Schiff

P.A.S. Periodic Acid Schiff Hotchkiss - Mc Manus	04-130802
PRODUCT AND APPLICATION	CODE

Minimum number of tests that can 100

be performed	
Completion time	50 minutes
Shelf life	1 year
Storage conditions	2-8°C
Additional equipment	Not required

#### Application

For highlighting normal or pathological tissue components, distinguished by adjacent glycol or amino hydroxyl groups on histological sections and on blood smears and cytology smears.

#### Method

•

Results

METHOD FOR HISTOLOGICAL SECTIONS

- 1) Bring the section to the distilled water.
- 2) Dispense 10 drops of reagent A onto the section: leave to act for 10 minutes.
- 3) Wash in distilled water.
- 4) Dispense 10 drops of reagent B onto the section: leave to act for 20 minutes.
- 5) Wash in distilled water.
- 6) Dispense 10 drops of solution C onto the section: leave to act for 2 minutes.
- 7) Drain the slide and, without washing, dispense 10 drops of reagent D onto the section: leave to act for 2 minutes.
- 8) Wash in distilled water.
- 9) Dispense 10 drops of reagent E onto the section: leave to act for 3 minutes.
- 10) Leave to develop in running water for 5 minutes.
  - 11) Dehydrate in the ascending series of alcohols, xylene and balsam.

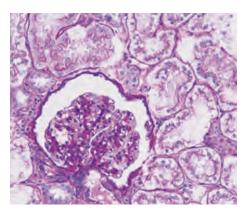
#### METHOD FOR BLOOD SMEARS AND CYTOLOGY SMEARS

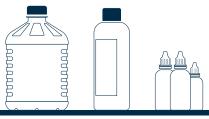
- 1) Place the air-dried smears in distilled water.
- 2) Dispense 10 drops of reagent A onto the smear: leave to act for 10 minutes.
- 3) Wash in distilled water.
- 4) Dispense 10 drops of reagent B onto the smear: leave to act for 20 minutes.
- 5) Wash in distilled water.
- 6) Dispense 10 drops of solution C onto the smear: leave to act for 2 minutes.
- 7) Drain the slide and, without washing, dispense 10 drops of reagent D onto the smear: leave to act for 2 minutes.
- 8) Wash in distilled water.
- 9) Dispense 10 drops of reagent E onto the smear: leave to act for 3 minutes.
- 10) Leave to develop in running water for 5 minutes.
- 11) Dehydrate in the ascending series of alcohols, xylene and balsam.

#### Result

PAS-positive substances	magenta red
Nuclei	blue

KIDNEY





PRODUCT AND APPLICATION		CODE
P.A.S A Periodic Acid Schiff - Amy	lase	04-130803
Minimum number of tests that can be performed	100	
Completion time	60 minutes	

Completion time	60 minutes
Shelf life	1 year
Storage conditions	2-8°C
Additional equipment	Not required

#### Application

Digestion on a histological section with an amylase solution followed by PAS reaction is indicated when you want to remove the glycogen so as to observe only the neutral epithelial mucins.

PAS/amylase reaction is the method of choice for evaluating the presence of glycogen in liver tissue on sections fixed in formalin and embedded in paraffin, and in muscle tissue on cryostat sections.

In both cases, the examination of adjacent sections, one of which has been treated with amylase, allows qualitative evaluation of the presence of glycogen.

#### Method

- 1) Bring the section to the distilled water.
- 2) Bring reagent A to room temperature.
- 3) Dispense 10 drops of reagent A: leave to act for 10 minutes at room temperature.
- 4) Wash the slide several times in distilled water.
- 5) Dispense 10 drops of reagent B onto the section: leave to act for 10 minutes.
- 6) Wash in distilled water.
- 7) Dispense 10 drops of reagent C onto the section: leave to act for 20 minutes.
- 8) Wash in distilled water.
- 9) Dispense 10 drops of solution D onto the section: leave to act for 2 minutes.
- 10) Drain the slide and, without washing, dispense 10 drops of reagent E onto the section: leave to act for 2 minutes.
- 11) Wash in distilled water.
- 12) Dispense 10 drops of reagent F onto the section: leave to act for 3 minutes.
- 13) Leave to develop in running water for 5 minutes.
- 14) Dehydrate in the ascending series of alcohols, xylene and balsam.

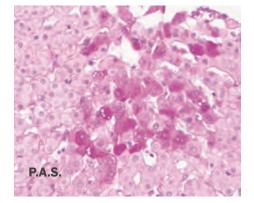
#### Result

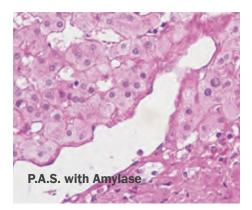
PAS-positive substances	magenta red
Nuclei	blue

Results

P.A.S. - A

LIVER





Perls

	PRODUCT AND APPLICATION	CODE	
•	Perls method for ferric iron	04-180807	
	Minimum number of tests that can be performed	72	
	Completion time	35 minutes	
	Shelf life	2 years	
	Storage conditions	15-25°C	
	Additional equipment	50 ml vertical histology jar, graduated cylinder ar	nd glass
		rod	

#### Application

Method indicated for viewing reactive ferric iron on tissue sections, blood smears and bone marrow smears.

Specificity - the Perls reaction does not show all the iron present in the tissue: iron bound to hemoglobin, malaria pigment, ferritin, pigments deriving from the use of acid formalin and ferrous iron does not react.

#### Method

METHOD FOR HISTOLOGICAL SECTIONS

- 1) Bring the section to the distilled water.
- 2) Transfer the entire contents of bottle A to a 50 ml Coplin jar. Add, in order, 30 ml of distilled water and 4 ml of reagent B. Stir briefly. Immerse the section for 20 minutes.
- 3) Wash thoroughly in distilled water.
- 4) Dispense 10 drops of reagent C onto the section: leave to act for 5 minutes.
- 5) Wash in distilled water.
- 6) Dehydrate by means of the ascending series of alcohols; xylene and balsam

METHOD FOR BLOOD SMEARS AND BONE MARROW SMEARS

- 1) Fix the previously dried smears in methanol for 3 minutes. Remove the slide and leave to dry.
- 2) Transfer the entire contents of bottle A to a 50 ml Coplin jar. Add, in order, 30 ml of distilled water and 4 ml of reagent B. Stir briefly. Immerse the section for 20 minutes.
- 3) Wash thoroughly in distilled water.
- 4) Dispense 10 drops of reagent C onto the smears: leave to act for 5 minutes.
- 5) Wash in distilled water.
- 6) Dry in air.

#### Result

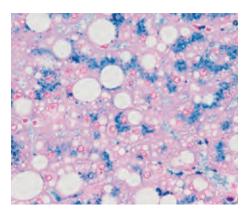
Reactive ferric iron	blue	
Nuclei	red	

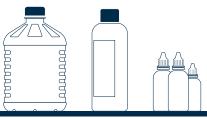
#### Notes:

Results

False positives may be caused by three easily identifiable factors:

- ferrocyanide-hydrochloric acid solution not freshly prepared;
- ferric ions contaminating the glassware and section stretching water (rust), use of metal instruments in contact with the solution (forceps etc.);
- asbestosis: asbestos, if present, can generate a positive reaction.





Perls - Van Gieson

#### **Staining and mounting**

	PRODUCT AND APPLICATION		CODE
1	Perls - Van Gieson method for ferrie	c iron and connective tissue	04-181807
	Minimum number of tests that can be performed	72	
	Completion time	35 minutes	
	Shelf life	2 years	
	Storage conditions	15-25°C	
	Additional equipment	50 ml vertical histology jar, gradua rod	ted cylinder and glass

#### Application

Method indicated for simultaneous highlighting of reactive ferric iron, collagen and connective tissue on tissue sections.

#### Method

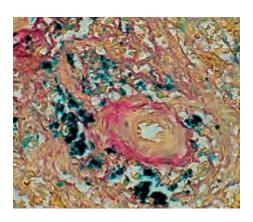
- 1) Bring the section to the distilled water.
- 2) Transfer the entire contents of bottle A to a 50 ml Coplin jar. Add, in order, 30 ml of distilled water and 4 ml of reagent B. Stir briefly. Immerse the section for 20 minutes.
- 3) Wash thoroughly in distilled water.
- 4) Dispense 10 drops of reagent C onto the section: leave to act for 10 minutes.
- 5) Wash in distilled water.
- 6) Dehydrate rapidly in the ascending series of alcohols, stopping for 1 minute in the last absolute; xylene and balsam.

Result	
Reactive ferric iron	blue
Collagen	purple red
Cytoplasm, muscle, stratum cor- neum of the epithelium, glia and erythrocytes	yellow

#### Notes:

False positives may be caused by three easily identifiable factors:

- ferrocyanide-hydrochloric acid solution not freshly prepared;
- ferric ions contaminating the glassware and section stretching water (rust), use of metal instruments in contact with the solution (forceps etc.);
- asbestosis: asbestos, if present, can generate a positive reaction.



**Results** 



#### **Picro Mallory Trichrome**

PRODUCT AND APPLICATION		CODE
Picro Mallory Trichrome		04-021822
Minimum number of tests that can be performed	100	
Completion time	40 minutes	
Shelf life	2 years	
Storage conditions	15-25°C	

Not required

#### **Application**

Trichrome stain recommended for connective sections.

#### Method

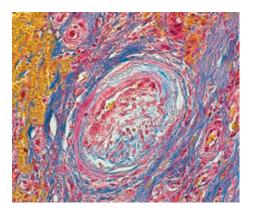
- 1) Bring section to distilled water.
- 2) Put on the section 5 drops of reagent A and 5 drops of reagent B: leave to act for 10 minutes.
- 3) Rinse in distilled water.

Additional equipment

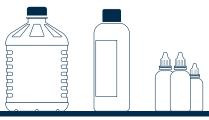
- 4) Blue 10 minutes in running tap water.
- 5) Put on the section 10 drops of reagent C: leave to act 2 minutes.
- 6) Rinse in distilled water.
- 7) Put on the section 10 drops of reagent D: leave to act 1 minute.
- 8) Rinse in distilled water.
- 9) Put on the section 10 drops of reagent E: leave to act 15 minutes.
- 10) Rinse in distilled water.
- 11) Put on the section 10 drops of reagent F: leave to act 1 minute.
- 12) Dehydrate rapidly through ascending alcohols, stop for 1 minute at the last absolute ethanol. Clear in xylene and mount.

#### CONNECTIVE TISSUE

Results



Nuclei:	dark brown
Collagen fibres:	dark blue
Ground substance of cartilage,	shades of blue
bone, mucus, basophil granules of	
hypophysis and amyloid:	
Neuroglia, axis cylinders and fibrin:	red
Acidophil granules of hypophysis:	orange
Myelin and erythrocytes:	Yellow
Elastic fibres:	pale pink to yellow



PRODUCT AND APPLICATION		CODE	P.T.A.H. Phosphotungstic Acid Hematoxylin
P.T.A.H. Phosphotungstic Acid Her	natoxylin	04-060802	
Minimum number of tests that can	100		
be performed			
Completion time	12 minutos + ovornight		

Completion time	13 minutes + overnight	
Shelf life	2 years	
Storage conditions	15-25°C	
Additional equipment	Not required	

#### Application

The method, originally proposed for staining the glia, is now mainly indicated for differentiating smooth muscle from striated muscle (by staining the isotropic bands of myofibrils of the skeletal muscle); it is also one of the methods of choice for fibrin.

#### Method

- 1) Bring the section to the distilled water.
- 2) Dispense 5 drops of solution A onto the section and add 5 drops of solution B: leave to act for 5 minutes.
- 3) Wash in distilled water.
- 4) Dispense 10 drops of solution C onto the section: leave to act for 5 minutes.
- 5) Wash in distilled water.
- 6) Pour the entire contents of the bottle of reagent D into the empty container attached to the pack, immerse the section in it and leave overnight.
- 7) Wash quickly in distilled water (3-4 seconds).
- 8) Dehydrate the section rapidly in the ascending series of alcohols, stopping for 1 minute in the last absolute; xylene and balsam.

#### Result

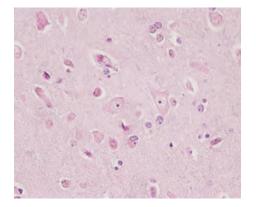
Nuclei, fibrin (most), myofibrils, astrocytes, certain elastic fibers, glia, myelin fibers Collagen, bone matrix, cartilage

dark blue

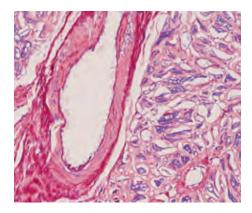
brick red in various shades

#### Results

BRAIN



BLOOD VESSEL





#### **Rapid frozen sections**

PRODUCT AND APPLICATION		CODE
Rapid frozen sections H&E staining	g kit	04-061010
Minimum number of tests that can be performed	100	
Completion time	Approximately 3 minutes	
Shelf life	2 years	
Storage conditions	15-25°C	

100 ml jar for buffer preparation, jar for washing

#### **Application**

Additional equipment

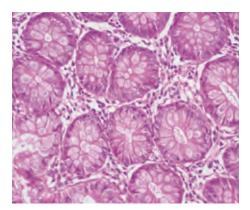
Rapid method for staining cryostat sections with a thickness of 6 microns.

#### Method

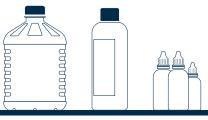
- 1) Preparation of the developing solution: dispense 10 drops of reagent B into a 100 ml jar. The kit is sufficient for the preparation of 100 developing solutions, we therefore recommended that you change the work solution frequently.
- 2) Place the section in the container labeled REAGENT A for 45 60 seconds.
- 3) Wash in tap water, 5 immersions.
- 4) Place in the developing solution, 5 immersions.
- 5) Wash in tap water, 5 immersions.
- 6) Place the section in the container labeled REAGENT C for 30 seconds.
- 7) 95° ethanol, 5 immersions.
- 8) 95° ethanol, 5 immersions.
- 9) Absolute ethanol, 5 immersions.
- 10) Absolute ethanol, 5 immersions.
- 11) Xylene, Bio-Clear or X-Free, 10 immersions.
- 12) Xylene, Bio-Clear or X-Free, 10 immersions.

#### COLON

**Results** 



Cytoplasm, connective tissue	pink in various shades and intensities
Nuclei	blue



PRODUCT AND APPLICATION		CODE	Congo Red
Highman's Congo red		04-210822	_
Minimum number of tests that can be performed	100		
Completion time	35 minutes		_
Shelf life	2 years		_
Storage conditions	15-25°C		_
Additional equipment	Not required		_

#### Application

Method for highlighting amyloid on tissue sections.

#### Method

Result

Nuclei

Amyloid substance

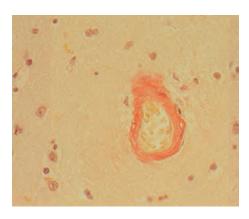
- 1) Bring the section to the distilled water.
- 2) Dispense 10 drops of reagent A onto the section: leave to act for 15 minutes.
- 3) Wash in distilled water.
- 4) Dispense 10 drops of reagent B onto the section: leave to act for 30 seconds.
- 5) Wash in running tap water for 5 minutes.
- 6) Dispense 10 drops of reagent C onto the section: leave to act for 2 minutes.
- 7) Dispense 10 drops of reagent D onto the section: leave to act for 5 minutes.
- 8) Leave to develop in tap water for 5 minutes.
- 9) Dehydrate by means of the ascending series of alcohols; xylene and balsam.

blue

brick red and birefringent in polarized light

#### Results

#### BLOOD VESSEL



#### **Sirius Red**

 PRODUCT AND APPLICATION
 CODE

 Sirius Red
 04-210923

 Minimum number of tests that can
 100

 be performed
 1 hour 15 minutes

 Completion time
 1 hour 15 minutes

 Shelf life
 2 years

15-25°C

Not required

#### **Application**

Storage conditions Additional equipment

Method for highlighting amyloid in tissues fixed in formalin and embedded in paraffin.

#### Method

Result

Nuclei

Amyloid substance

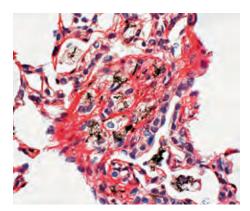
- 1) Bring the section to the distilled water.
- 2) Prepare the humid chamber and place the slide in it with the section facing up. Dispense 10 drops of reagent A onto the section, close the humid chamber and incubate in an oven at 60°C. Leave to act for 60-90 minutes.
- 3) Dispense 10 drops of reagent B onto the section for 1 2 minutes.
- 4) Drain the slide and dispense 10 drops of reagent C onto the section: leave to act for 1 - 2 minutes.
- 5) Dispense 10 drops of reagent D onto the section. Leave to act for 5 minutes.

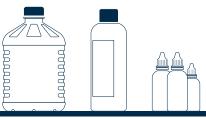
pink - red

blue

- 6) Leave to develop in running water for 5 minutes.
- 7) Dehydrate in the ascending series of alcohols, xylene and balsam.

#### AMYLOID SUBSTANCE





**Picrosirius Red** 

#### **Staining and mounting**

PRODUCT AND APPLICATION		CODE	
Picrosirius Red		04-121873	
Minimum number of tests that can be performed	100		
Completion time	60 minutes		
Shelf life	2 years		
Storage conditions	15-25°C		
Additional equipment	Not required		

#### Application

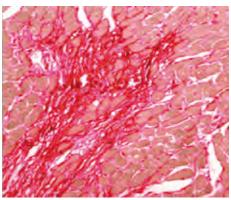
Method indicated for highlighting collagen fibers and bile pigments on tissue sections fixed in formalin and embedded in paraffin.

#### Method

- 1) Bring the section to the distilled water. •
- 2) Dispense 10 drops of reagent A onto the section: leave to act for 50 minutes.
- 3) Wash briefly in distilled water.
- 4) Dispense 10 drops of reagent B onto the section: leave to act for 2 minutes. Repeat twice.
- 5) Wash briefly in distilled water and drain the slide.
- 6) Dispense 10 drops of reagent C onto the section: leave to act for 3 minutes.
- 7) Leave to develop in tap water: 3 minutes.
- 8) Wash in distilled water and dehydrate rapidly by means of the ascending series of alcohols, leaving for 1 minute in the last absolute: xylene and balsam.

Result		
Bilirubin	green	
Collagen fibers	red	
Nuclei	blue	
Erythrocytes	red	

## **Results**





#### **Methenamine Silver**

Methenamine Silver P.A.S.M.	04-043822
PRODUCT AND APPLICATION	CODE

Minimum number of tests that can 100 be performed

Completion time	1 hour 15 minutes
Shelf life	1 year
Storage conditions	2-8°C
Additional equipment	Oven

#### Application

Method used for viewing argyrophilic elements and mucopolysaccharides (basal membranes, mycetes, bacteria, etc.) on tissue sections. It is the method of choice for studying the basal membrane in renal biopsy.

#### Method

- 1) Bring the sections to the distilled water.
- 2) Dispense 10 drops of reagent A onto the section: leave to act for 30 minutes.
- 3) Wash in distilled water.
- 4) Prepare the humid chamber and place the slide in it with the section facing up. Dispense 10 drops of reagent B into the small dish attached to the pack, add 10 drops of reagent C and 10 drops of reagent D, stir and place the solution thus obtained on the section: close the humid chamber and incubate in an oven at 60 °C. Leave to act for 30-40 minutes.
- 5) Remove the humid chamber from the oven, open the lid and check the tone of the impregnation: if the blackening is correct, leave the slide to cool for 5 minutes and then wash it in distilled water; if it is insufficient, incubate in the oven again and check every 5 minutes.
- 6) Dispense 10 drops of reagent E onto the section: leave to act for 1 minute.
- 7) Wash in distilled water.
- 8) Dispense 10 drops of reagent F onto the section: leave to act for 1 minute.
- 9) Wash in distilled water.
- 10) Dehydrate by means of the ascending series of alcohols, xylene and balsam.

#### Result

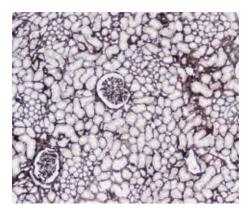
Results

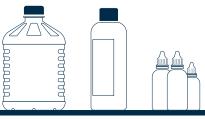
Basal membranes, glycogen, dish black of mycetes and bacteria

#### WARNINGS

As for all reactions involving silver salts, it is essential to use rigorously clean glassware and good-quality distilled or deionized water. Furthermore, do not bring metal instruments (forceps, etc.) into contact with reagents containing silver salts.

**KIDNEY** 





	PRODUCT AND APPLICATION		CODE	Van Gieson Trichrome
•	Van Gieson Trichrome		04-030802	-
	Minimum number of tests that can be performed	100		
	Completion time	35 minutes		-
	Shelf life	2 years		-
	Storage conditions	15-25°C		-
	Additional equipment	Not required		-

#### Application

Method of choice for connective tissue, particularly indicated for highlighting collagen fibers and differentiating them from connective tissue.

#### Method

Result Nuclei

Collagen fibers

Cytoplasm, smooth and striated

muscle, stratum corneum of the epithelium, glia and erythrocytes

- 1) Bring the section to the distilled water.
- 2) Dispense 5 drops of reagent A onto the section and add 5 drops of reagent B : leave to act for 10 minutes.
- 3) Leave to develop in tap water for 10 minutes.
- 4) Dispense 10 drops of solution C onto the section: leave to act for 10 minutes.
- 5) Wash quickly (2-3 seconds) in distilled water and dehydrate rapidly in the ascending
- series of alcohols, stopping for 1 minute in the last absolute; xylene and balsam.

black

yellow

purple red

#### Results

#### CONNECTIVE TISSUE



#### Verhoeff

#### Application

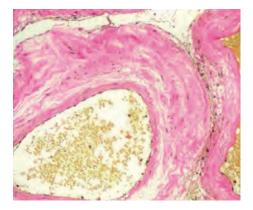
Method for demonstrating elastic fibers on histological sections, particularly indicated for vascular pathology.

#### Method

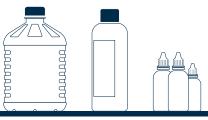
- 1) Bring the section to the distilled water.
- 2) Dispense 10 drops of reagent A onto the section: leave to act for 30 minutes.
- 3) Wash in distilled water.
- 4) Differentiate in tap water.
- 5) Place the slide in the humid chamber and dispense 8 drops of reagent B + 4 drops of reagent C + 4 drops of reagent D onto the section. Leave to act for 25 minutes.
- 6) Wash in distilled water.
- 7) Differentiate with reagent E: 2 or 3 changes of 15 seconds each.
- 8) Wash thoroughly in distilled water.
- 9) Dispense 10 drops of reagent F onto the section: leave to act for 1 minute.
- 10) Wash in distilled water.
- 11) Dehydrate by means of the ascending series of alcohols; xylene and balsam.

#### Results

ARTERY



Elastic fibers and nuclei	black
Collagen	red
Other tissue elements	yellow



Von Kossa

#### **Staining and mounting**

PRODUCT AND APPLICATION		CODE
Von Kossa method for calcium		04-170801
Minimum number of tests that can be performed	100	
Completion time	1 hour 25 minutes	
Shelf life	1 year	

#### **Application**

Storage conditions

Additional equipment

Method indicated for viewing calcium ions on histological sections.

#### Method

- 1) Bring the section to the distilled water.
- 2) Dispense 10 drops of reagent A onto the section: leave to act for 10 minutes.

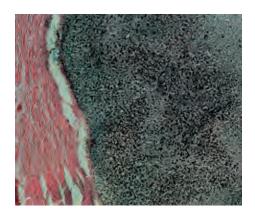
2-8°C

Not required

- 3) Wash thoroughly in distilled water.
- 4) Dispense 10 drops of reagent B onto the section: leave to act in the dark for 1 minutes.
- 5) Wash thoroughly in distilled water.
- 6) Dispense 10 drops of distilled water onto the section and add 10 drops of reagent
- C: leave to act for 5 minutes (until the silver salts turn black).
- 7) Wash in distilled water.
- 8) Dispense 10 drops of reagent D onto the section: leave to act for 5 minutes.
- 9) Wash in distilled water.
- 10) Dispense 10 drops of reagent E onto the section: leave to act for 5 minutes.
- 11) Wash in distilled water and dehydrate in the ascending series of alcohols; xylene and balsam.

Result	
Sites where calcium salts were	black
present	
Nuclei	red

#### BONE



#### Warthin-Starry

PRODUCT AND APPLICATION

Warthin-Starry method for spirochetes 04-040903

CODE

 Minimum number of tests that can
 40

 be performed
 Completion time

 Completion time
 1 hour 45 minutes

 Shelf life
 1 year

 Storage conditions
 2-8 ° C

 Additional equipment
 Oven, jar for buffer dilution, graduated pipette, glass rod

#### Application

Method for highlighting spirochetes.

#### Method

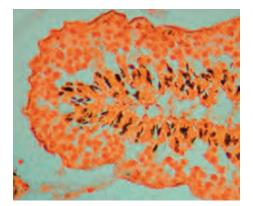
- 1) Bring the section to the distilled water.
- 2) Prepare the impregnating solution: pour 13 ml of distilled water into the container, then add 4.5 ml of reagent A and 20 drops of reagent B. Stir briefly with a glass rod previously washed in distilled water.
- 3) Place the section in the solution and incubate for 90 minutes at 60-70°C.
- 4) Remove the container from the oven and leave to cool for 5 minutes.
- 5) While the impregnation reaction takes place, prepare the developing solution. Note: you are advised to carry out the indicated operations during the last 12 minutes of incubation started in step 3. Preheat one bottle C and one bottle D in an oven at 50 °C for 10 minutes.

Pour the entire contents of the two preheated bottles into the second container available for slides (beware of the temperature of the bottles – use protective gloves), stir briefly with a glass rod previously washed in distilled water and then pour in the entire contents of one bottle E and stir again.

- 6) Place the section in the developing solution you have just prepared and place in an oven at 50 ° C for 5 10 minutes.
- 7) Wash in hot running water for 2 minutes.
- 8) Dehydrate by means of the ascending series of alcohols, xylene and balsam.

#### Results

SPIROCHETES

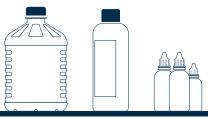


#### Result

Spirochetes and other micro- organisms	black
Background	golden brown

#### WARNINGS

- For washing, it is imperative to use top-quality distilled water.
- Do not use Poly-L-Lysine coated slides.
- Do not use metal objects (racks, forceps).



PRODUCT AND APPLICATION		CODE		Weigert - long method
	Weigert for elastic fibers (long me	thod)	04-050802	
	Minimum number of tests that can be performed	100		
	Completion time	Overnight + 25 minutes		
	Shelf life	2 years		
	Storage conditions	15-25°C		

#### **Application**

Additional equipment

Method indicated for demonstrating elastic fibers on histological sections.

#### Method

- 1) Bring the section to the distilled water.
- 2) Dispense 5 drops of solution A onto the section and add 5 drops of solution B: leave to act for 5 minutes.

Histology jar with lid

- 3) Wash in distilled water.
- 4) Dispense 10 drops of solution C onto the section: leave to act for 5 minutes.
- 5) Wash in distilled water.
- 6) Pour the reagent D into a vertical histology jar, immerse the section in it and close firmly: leave to act overnight.

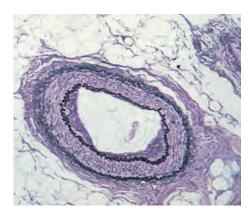
After use, in order to minimize evaporation of the ethanol, you are advised to return the solution to its original bottle.

- 7) Wash in distilled water.
- 8) Dispense 10 drops of solution E onto the section: leave to act for 10 minutes.
- 9) Wash in distilled water.
- 10) Dehydrate by means of the ascending series of alcohols; xylene and balsam.

Result Elastic fibers

from dark blue to black







#### Weigert - rapid method

PRODUCT AND APPLICATION		CODE
Weigert for elastic fibers (rapid me	thod)	04-052812
Minimum number of tests that can be performed	100	
Completion time	60 minutes	
Shelf life	2 years	
Storage conditions	15-25°C	

Graduated pipette

#### **Application**

Method indicated for demonstrating elastic fibers on histological sections, particularly indicated for vascular pathology.

#### Method

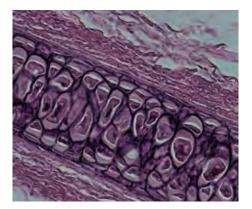
- 1) Bring the section to the distilled water.
- 2) Dispense 10 drops of solution A onto the section: leave to act for 5 minutes.
- 3) Wash in distilled water.

Additional equipment

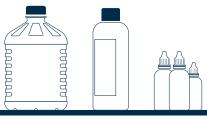
- 4) Prepare the humid chamber as follows: soak the disc of filter paper with 20 drops of reagent B, insert the slide in the humid chamber and dispense 10 drops of reagent C onto the section. Close the lid and incubate for 30 minutes.
- 5) Wash in distilled water.
- 6) Dispense 10 drops of reagent D onto the section: leave to act for 2 minutes.
- 7) Wash in running water for 5 minutes
- 8) Wash in distilled water.
- 9) Dispense 10 drops of reagent E onto the section: leave to act for 5 minutes.
- 10) Wash in distilled water.
- 11) Dehydrate by means of the ascending series of alcohols; xylene and balsam.

#### Results

ELASTIC FIBERS



Elastic fibers	purple - brown
Nuclei	red



	PRODUCT AND APPLICATION		CODE	Weigert Van Gieson long method
•	Weigert Van Gieson for elastic fibers	and connective tissue(long method)	04-051802	
	Minimum number of tests that can	100		
	be performed			
	Completion time	50 minutes + overnight		
	Shelf life	2 years		
	Storage conditions	15-25°C		

Vertical histology jar with lid

#### Application

Additional equipment

Combined method for viewing elastic fibers, connective tissue, collagen and nuclei on the same preparation.

Van Gieson trichrome is the most commonly used method in association with Weigert staining for elastic fibers.

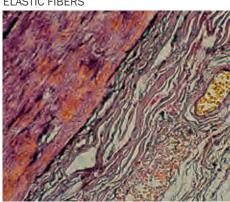
#### Method

- 1) Bring the section to the distilled water.
- 2) Dispense 10 drops of solution A onto the section: leave to act for 5 minutes.
- 3) Wash in distilled water.
- 4) Pour the reagent B into a vertical histology jar, immerse the section in it and close firmly: leave to act overnight. After use, you are advised to return the solution to its original bottle.
- 5) Wash in distilled water.
- 6) Dispense 10 drops of reagent C onto the section: leave to act for 10 minutes.
- 7) Wash in distilled water.
- 8) Dispense 5 drops of solution D onto the section and add 5 drops of solution E: leave to act for 10 minutes.
- 9) Leave to develop in tap water for 10 minutes.
- 10) Dispense 10 drops of solution F onto the section: leave to act for 7 minutes.
- 11) Wash quickly (2-3 seconds) in distilled water and dehydrate rapidly in the ascending series of alcohols, stopping for 1 minute in the last absolute; xylene and balsam.

purple - brown
black
red, in various shades
yellow

CONNECTIVE TISSUE AND ELASTIC FIBERS

Results



p\_**105**/139

#### Weigert Van Gieson rapid method

PRODUCT AND APPLICATION

CODE

• Weigert Van Gieson for elastic fibers and connective tissue(rapid method) 04-053812

Minimum number of tests that can	100
be performed	
Completion time	1 hour 20 minutes
Shelf life	2 years
Storage conditions	15-25°C
Additional equipment	Graduated pipette

#### Application

Combined method for viewing elastic fibers, connective tissue, collagen and nuclei on the same preparation.

Van Gieson trichrome is the most commonly used method in association with Weigert staining for elastic fibers.

#### Method

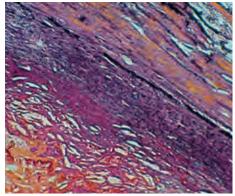
- 1) Bring the section to the distilled water.
- 2) Dispense 10 drops of solution A onto the section: leave to act for 5 minutes.
- 3) Wash in distilled water.
- 4) Prepare the humid chamber as follows: soak the disc of filter paper with 20 drops of reagent B, insert the slide in the humid chamber and dispense 10 drops of reagent C onto the section. Close the lid and incubate for 30 minutes.
- 5) Wash in distilled water.
- 6) Dispense 10 drops of reagent D onto the section: leave to act for 2 minutes.
- 7) Wash in tap water for 5 minutes.
- 8) Wash in distilled water.
- 9) Dispense 5 drops of solution E onto the section and add 5 drops of solution F: leave to act for 10 minutes.
- 10) Leave to develop in tap water for 10 minutes.
- 11) Dispense 10 drops of solution G onto the section: leave to act for 10 minutes.
- 12) Wash quickly (2-3 seconds) in distilled water and dehydrate rapidly in the ascending series of alcohols, stopping for 1 minute in the last absolute; xylene and balsam.

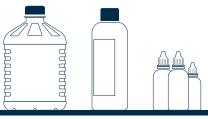
#### Result

Elastic fibers	purple - brown
Nuclei	black
Collagen	red, in various shades
Connective tissue, erythrocytes	yellow

Results

CONNECTIVE TISSUE AND ELASTIC FIBERS





PRODUCT AND APPLICATION		CODE	
Ziehl-Neelsen for mycobacteria		04-110802	
Minimum number of tests that can be performed	100		
Completion time	50 minutes		
Shelf life	2 years		
Storage conditions	15-25°C		

#### Application

For highlighting pathogenic mycobacteria with particular regard to Koch's bacillus, on histological sections, sputum and culture smears, and appositions.

Not required

#### Method

- 1) Bring the sections to the distilled water.
- 2) Dispense 10 drops of reagent A onto the section: leave to act for 10 minutes.
- 3) Wash in distilled water.

Additional equipment

- 4) Dispense 10 drops of reagent B onto the section: leave to act for 30 minutes.
- 5) Wash in distilled water and dry the slide with filter paper.
- 6) Dispense 10 drops of reagent C onto the section: leave to act for 1 minute.
- 7) Wash in tap water for 3 minutes.
- 8) Dispense 10 drops of reagent D onto the section: leave to act for 2 minutes.
- 9) Wash in distilled water, develop for 5 minutes in running water.
- 10) Dehydrate by means of the ascending series of alcohols, xylene and balsam.

#### Result

Koch's bacillus and other acid-resistant elements

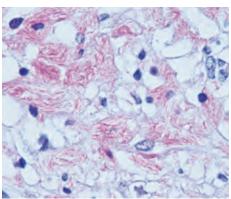
Nuclei

blue - violet

red

### Ziehl-Neelsen

LUNG



#### **Ziehl-Neelsen Fite**

	PRODUCT AND APPLICATION		CODE
•	Ziehl-Neelsen Fite for mycobacteri	а	04-111802
	Minimum number of tests that can be performed	100	
	Completion time	45 minutes	
	Shelf life	2 years	
	Storage conditions	15-25°C	
	Additional equipment	Not required	

#### Application

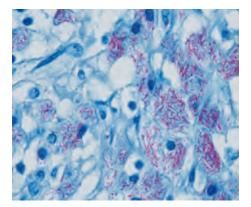
For highlighting pathogenic mycobacteria with particular regard to Koch's and Hansen's bacillus, on histological sections, sputum and culture smears, and appositions.

#### Method

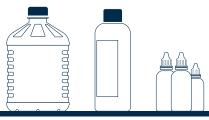
- 1) Bring the sections to the distilled water.
- 2) Dispense 10 drops of reagent A onto the section: leave to act for 10 minutes.
- 3) Wash in distilled water.
- 4) Dispense 10 drops of reagent B onto the section: leave to act for 30 minutes.
- 5) Wash in distilled water and dry the slide with filter paper.
- 6) Dispense 10 drops of reagent C onto the section: leave to act for 1 minute.
- 7) Wash in tap water for 3 minutes.
- 8) Dispense 10 drops of reagent D onto the section: leave to act for 1 minute.
- 9) Wash in distilled water.
- 10) Dehydrate by means of the ascending series of alcohols, xylene and mounting medium.

LUNG

Results



Koch's bacillus, Hansen's bacillus	red – violet
and other acid-resistant elements	
Background contrast	light blue



#### Kits and solutions for enzyme histochemistry

Microscopic examination of sections of muscle biopsies is an essential tool in the diagnosis of neuromuscular disorders.

Any laboratory choosing to conduct histo-enzymatic tests on muscle biopsies encounters a series of problems:

- high toxicity of certain reagents
- solutions that are difficult and complex to standardize
- storage of solutions at -20°C
- poor reproducibility of final results.

To overcome all these problems, Bio-Optica has developed ready-to-use kits for enzyme histochemistry. Enzyme histochemistry kits eliminate the difficulties and risks associated with the preparation of stain solutions, thus ensuring reproducible results.

PRODUCT AND APPLICATION

CODE

 ATPase
 30-30125LY

 Method of choice for determining the types of muscle fibers. For use with cryostat sections of striated muscle with a thickness of 8 µm.

 The solutions, supplied in ready-to-use form, make it possible to perform the method on three serial sections of the sample simultaneously.

 For correct application of the method, it is necessary to use reagents that have been brought to room temperature.

•

Result		
Nuclei	blue	
Section 10.4 - preincubation at pH	10.4	
Type 1 fibers	white - beige	
Type 2A fibers	brown - black	
Type 2B fibers	brown - black	
Section 4.7 - preincubation at pH 4.7		
Type 1 fibers	brown	
Type 2A fibers	white - beige	
Type 2B fibers	brown - dark brown	
Section 4.3 - preincubation at pH 4.3		
Type 1 fibers	brown	
Type 2A fibers	white - beige	
Type 2B fibers	beige	



**ATPase** 

## Bio - Optica

## Cytochrome C oxidase



PRODUCT AND APPLICATION
Cytochrome C oxidase

CODE

CODE

30-30122LY

30-30115LY

Evaluation of Cytochrome C oxidase activity.

#### Result

Activity of Cytochrome C oxidase beige positive

## Non-specific esterase



## Non-specific esterase

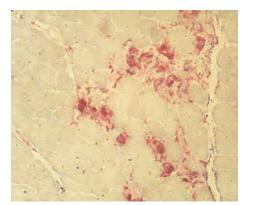
PRODUCT AND APPLICATION

Highlighting positive enzymatic activity of esterase in denervated fibers.

#### Result

Angular atrophic fibers	beige
Muscle plaques	brown
Lipofuscins	brown
Lysosomal activity	brown

## Acid phosphatase



 PRODUCT AND APPLICATION
 CODE

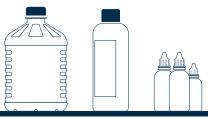
 Acid phosphatase
 30-30118LY

 Highlighting enzymatic activity of acid phosphatase.
 Present in macrophages and lysosomes; identifies necrosis and regeneration.

 Result
 Positive enzymatic activity of acid red phosphatase

 Posphatase
 Vertical activity of acid red phosphatase

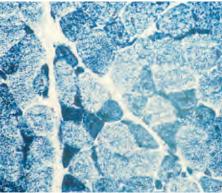
Background and nuclei green

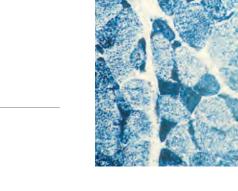


## **Staining and mounting**

PRODUCT AND APPLICATION		CODE	Alkaline phosphatase
Alkaline phosphatase		30-30121LY	1 and 1
Evaluating enzymatic activity of al	kaline phosphatase.		7 1 2
Useful for highlighting the sites of	phagocytosis and inflammation in		
muscle biopsies.			and the second
			· · · · ·
Result			
Positive enzymatic activity of	black		12 11 10
alkaline phosphatase			ALL AND
Background	yellow ochre		AN NY
			37

Phosphofructokinase (PFK)





PRODUCT AND APPLICATION

CODE

30-30123LY

CODE

30-30117LY

Phosphorylase

PRODUCT AND APPLICATION

metabolism of glycogen.

Positive PFK activity

Result

•

Phosphofructokinase (PFK)

Highlighting enzymes belonging to the phosphorylase class involved in glycogenolysis.

Highlighting enzymatic activity of phosphofructokinase (PFK), useful for determining whether glycogen storage disease depends on a deficiency of phosphofructokinase or on other enzymes involved in the

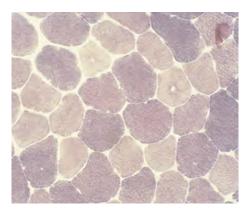
dark blue

#### Result

Phosphorylase enzyme activity

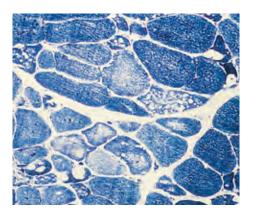
various shades of blue

### **Phosphorylase**



## Bio - Optica

## Myoadenylate deaminase



PRODUCT AND APPLICATION

 Myoadenylate deaminase
 30-30116LY

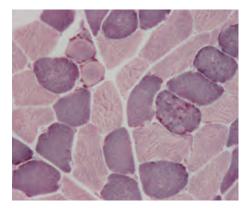
 Highlighting of enzymatic activity of myoadenylate deaminase
 (AMPDA).

CODE

#### Result

Positive enzymatic activity of blue myoadenylate deaminase

## NADH diaphorase



	PRODUCT AND APPLICATION	CODE
•	NADH diaphorase	30-30113LY
	Evaluation of NADH diaphorase activity. Staining useful for	
	distinguishing type 1 and type 2 muscle fibers, often used in	
	conjunction with ATPase evaluation.	

#### Result

### Succinate dehydrogenase



 PRODUCT AND APPLICATION
 CODE

 Succinate dehydrogenase
 30-30114LY

 Evaluation of enzymatic activity of succinate dehydrogenase (SDH) detected specifically in the mitochondria.
 Succinate dehydrogenase (SDH)

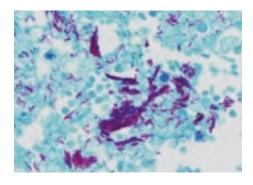
 Result
 Positive SDH activity
 gray – blue



## Staining and mounting

PRODUCT AND APPLICATION		CODE
Ziehl-Neelsen Fite for microbiolo	gy	04-111803
To show pathogenic mycobacteria	(especially Koch's bacillus) in	
sputum smears and culture smea	irs.	
Result		
Koch's bacillus and other acid	red	
resistant elements		
Background	blue	

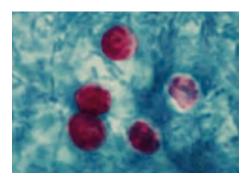
## Koch's bacilli



PRODUCT AND APPLICATION	CODE
Ziehl-Neelsen for Cryptosporidium	04-110803
To show Cryptosporidium sp. oocysts in fecal smears.	
Result	

Result		
Cryptosporidium sp. oocysts	red	
(4 - 6 μm diameter)		
Background	Pale green	

## Cryptosporidium

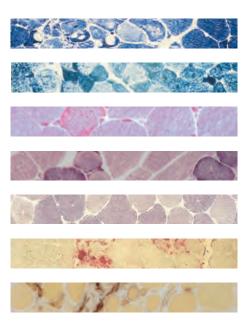


## Fixatives for enzyme histochemistry

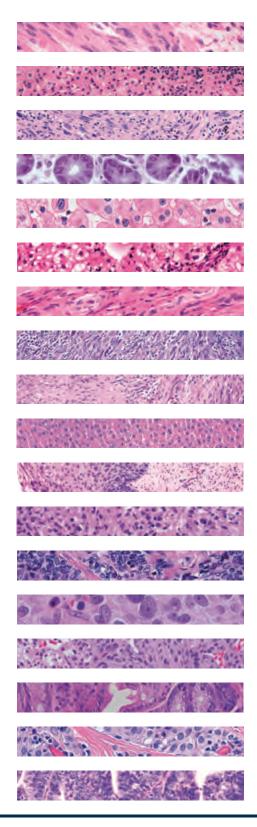
PRODUCT	PACK	DESCRIPTION	CODE
Backer fixative	1x500 ml	Facilitates staining with Oil Red O	30-30111
Fixative for acid phosphatase	1x100 ml	For use in enzyme histochemical staining for acid phosphatase	30-30120

## Staining solutions for enzyme histochemistry

PRODUCT	PACK	DESCRIPTION	CODE
Oil Red O solution Buffered methyl green solution	1x100 ml 1x100 ml	Specific stain for lipids Green nuclear stain	30-30112 30-30119
Gomori trichrome solution	1x100 ml	Stain for the morphological study of muscle fiber and connective tissue	30-30110
solution		,	



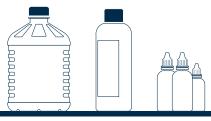
# Bio-Optica



## Hematoxylin

Bio-Optica provides operators with a complete range of nuclear stains; all solutions are stable and yield excellent cellular details.

PRODUCT AND DESCRIPTION	PACK	CODE
Mayer's hemalum	1x500 ml	05-M06002
Medium-intensity stain	1x1	05-06002/L
	1x2,5 I	05-06002E
Harris hematoxylin for histology	1x500 ml	05-M06004
Stain with a high concentration of hematoxylin	1x1	05-06004/L
	1x2,5 I	05-06004E
Carazzi's hemalum	1x500 ml	05-M06012
Lower concentration of hematoxylin	1x1	05-06012/L
Gill 1 hematoxylin	1x1	05-06013/L
Similar to Carazzi's hemalum		
Gill 2 hematoxylin	1x500 ml	05-M06014
Similar to Mayer's hemalum	1x1	05-06014/L
	1x2,5 I	05-06014E
Gill 3 hematoxylin	1x500 ml	05-M06015
Similar to Harris hematoxylin for histology	1x1	05-06015/L
	1x2,5 I	05-06015E
Weigert A ferric hematoxylin	1x150 ml	05-B06008/A
For trichrome staining	1x1	05-06008A/L
Weigert B ferric hematoxylin	1x150 ml	05-B06008/B
For trichrome staining	1x1	05-06008B/L
P.T.A.H. – Phosphotungstic Acid Hematoxylin	1x1	05-10017/L
For staining muscle fibers and nerves		

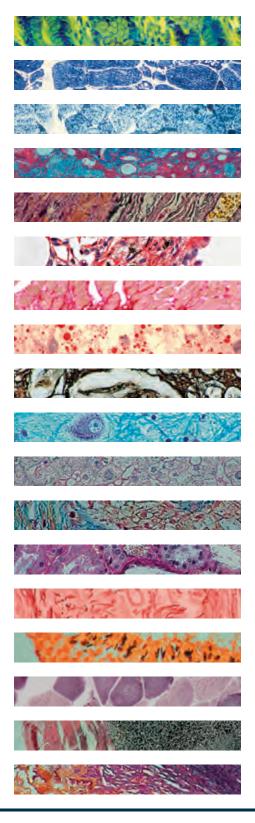


## **Staining and mounting**

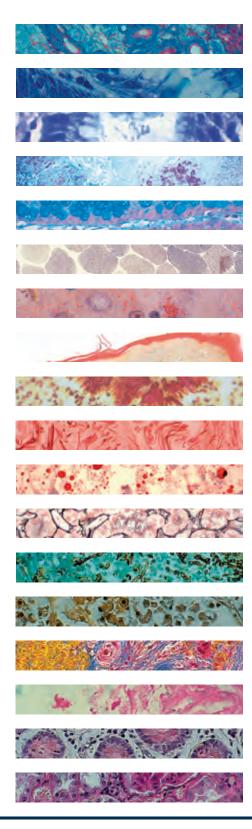
#### **Histology solutions**

In addition to a number of important functional benefits (safety, time saving, reduced workload, easy estimation of costs per test), Bio-Optica's ready-to-use solutions yield excellent, reproducible results. These are essential characteristics for meeting the requirements of laboratories adhering to high quality standards.

	PRODUCT AND DESCRIPTION	PACK	CODE
•	Alcian Blue pH 2,5 Mowry	1x500 ml 1x1 l	05-M26003 05-26003/L
•	Masson Aniline Blue	1x150 ml	05-B10006
•	Basic Fuchsin solution	1x1	05-07012/L
•	Cresyl Blue	1x150 ml	05-B14002
•	New Blue Methylene	1x150 ml	05-B14003
•	Toluidine Blue polychrome	1x150 ml	05-B23001
		1x500 ml	05-M23001
•	Mayer's Carmalum	1x150 ml	05-B07009
•	Crystal violet Metachromatic	1x150 ml	05-B31001
•	Eosin Y aqueous solution 1%	1x500 ml	05-M10007
		1x1	05-10007/L
		1x2,5	05-10007E
•	Eosin Y alcoholic solution 0,5%	1x500 ml	05-M10009
		1x1	05-10009/L
		1x2,5	05-10009E
•	Eosin Y Plus alcoholic solution	1x1	05-11007/L
•	Eosin Phloxin solution	1x500 ml	05-M10020
		1x1	05-10020/L
•	Ziehl carbol fuchsin	1x500 ml	05-M20007
•	Fuchsin ponceau Masson	1x150 ml	05-B10005
•	Giemsa Pappenhaim	1x500 ml	05-M12005
		1x1	05-12005/L
		1x2,5	05-12005E
•	Luxol fast blue Klüver Barrera	1x150 ml	05-B18001
٠	May Grunwald Pappenheim	1x500 ml	05-M12002
		1x1	05-12002/L
		1x2,5 I	05-12002E
•	Nuclear Fast Red	1x150 ml	05-B07006
		1x500 ml	05-M07006
•	Picrofuchsin Van Gieson	1x500 ml	05-M10012
٠	Picro Mallory – Acid Fuchsin	1x150 ml	05-B10014
٠	Schiff's reagent Feulgen	1x500 ml	05-M07007
٠	Schiff's reagent Hotchkiss McManus	1x500 ml	05-M20001
٠	Safranin solution	1x1	05-07008/L
٠	Sudan III Herxheimer	1x150 ml	05-B27001
٠	Sudan Black	1x150 ml	05-B27002
٠	Light green Goldner	1x500 ml	05-M10008

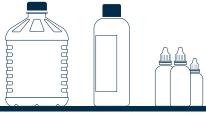






## Reagents

PRODUCT AND DESCRIPTION	PACK	CODE
Phosphomolybdic acid Masson	1x500 ml	05-M05003
Periodic acid 1%	1x500 ml	05-M05030
Picric acid aqueous solution 1.2%	1x500 ml	05-M05027
Scott's water	10x500 ml	05-M05023
Albumine Mallory	1x150 ml	05-B04002
Alcohol Borax Mowry	1x150 ml	05-B05011
Lithium Carbonate solution	1x500 ml	05-M05016
Lugol solution	1x500 ml	05-M05015



## **Staining and mounting**

### **Mount Quick Aqueous**

Synthetic mounting medium, dissolved in water. For use when dehydration causes loss of staining characteristics. Compatible with hematoxylin-eosin.

PACK	CODE
9x30 ml	05-1740

#### Immersion oil for microscopes

Type A oil for microscopes.

PACK	CODE
1x30 ml	08-1730/A30
9x30 ml	08-1730/A270

#### **BioMount HM**

Synthetic mounting medium, dissolved in xylene, particularly indicated for use with the automatic coverslipper.

PACK	CODE
1x100 ml	05-BMHM100
8x500 ml	05-BMHM508

## **CVR Mount**

Xylene based and isoparaffin based mounting medium, indicated for use with the automatic coverslipper.

PRODUCT	PACK	CODE	
CVR Mount	1x500 ml	05-CVR500	
CVR Mount Ultra (Isoparaffin)	1x500 ml	05-CVR501	

## Coverslips

Cleaned, degreased, high-quality coverglasses; free from dust, dirt and cracks.

DIMENSIONS	PACK	CODE
24 x 40 mm	1000 pcs.	09-2040
24 x 50 mm	1000 pcs.	09-2050
24 x 60 mm	1000 pcs.	09-2060
50 x 65 mm	1000 pcs.	09-5065

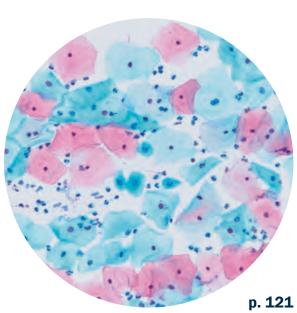




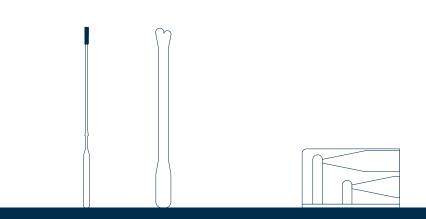




















## **Dual cyto cuvettes**

Centrifugation cuvettes with dual chamber.

PRODUCT	PACK	CODE
Medium absorption High absorption	500 pcs. 500 pcs.	14-080 14-070
Fixatives		
PRODUCT AND DESCRIPTION	PACK	CODE
• <b>Cy-Fix</b> Fixative for liquid-based cytology	54x25 ml	05-01C50P
• <b>Bio-fix</b> Spray fixative for vaginal cytology	4x200 ml	05-X200
• Saccomanno's Fixative Fixative for samples with mucus	1x1	05-01043/L



## **Bio-Agar for embedding cytological samples**

Aqueous gel for embedding cytological samples, encapsulates and retains cells during processing. Useful for processing centrifuged cells and fragile biopsies.

PRODUCT 15x10 ml

CODE

05-9803S



## Cytology

## Papanicolaou staining solutions

Fast staining, bright colors and excellent cellular details. The solutions are methanol-free.

PACK	CODE
1x500 ml	05-12011
1x1	05-12011/L
1x2,5 I	05-12011E
1x500 ml	05-12013
1x1	05-12013/L
1x2,5 I	05-12013E
1x500 ml	05-12019
1x1	05-12019/L
1x2,5 I	05-12019E
1x500 ml	05-12017
1x1	05-12017/L
	1x500 ml 1x1 l 1x2,5 l 1x500 ml 1x1 l 1x2,5 l 1x500 ml 1x1 l 1x2,5 l 1x500 ml



## May Grünwald Giemsa solutions

Ready-to-use solutions for the differentiation of cellular elements in blood smears, spleen tissue samples, lymph node tissue and bone marrow biopsies.

May Grunwald Giemsa for smears	1x1	04-080802/L
	1x2,5	05-12005E
1	1x1	05-12005/L
Giemsa	1x500 ml	05-M12005
	1x2,5	05-12002E
	1x1	05-12002/L
May Grünwald	1x500 ml	05-M12002
PRODUCI	PACK	CODE
PRODUCT	PACK	CODE



# Bio - Optica

## May Grünwald Giemsa kit

PRODUCT AND APPLICATION		CODE	
•	May Grünwald Giemsa kit for smea	ars 04-0808	02
	Minimum number of tests that can be performed	50 preparations	
	Completion time	35 minutes	
	Shelf life	2 years	
	Storage conditions	15-25°C	
	Additional equipment	1000 ml calibrated flask, 100 ml graduated 100 ml histology jar	cylinder,

#### **Application**

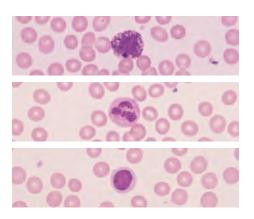
.

For differential staining of cellular elements in blood smears, and spleen, lymph node and bone marrow tissue samples.

#### Method

- 1) Pour 100 ml of reagent B (buffer concentrated solution) into a 1000 ml calibrated flask and top up to the required volume with tap water (buffer - work solution). Keep the two buffer solutions at 4° - 6° centigrade.
- 2) Dispense 10 drops of reagent A onto the slide; leave to act for 5 minutes. Note: Where considered appropriate, the above step can be performed by working in the jar without making any change to the times (in this case the reagent must be recovered).
- 3) Wash in running water for 1 minute.
- 4) Pour 10 ml of solution C into a cylinder containing 90 ml of buffer solution B (work solution), pour the mixture into a vertical histology jar and immerse the slide in it for 15 minutes.
- 5) Wash in running water for 1 2 minutes.
- 6) Wash the slide first in filter paper, then in the air for 5 minutes.

#### Results



#### Result

Robult	
Nuclei	violet red, pink
Basophilic cytoplasm	blue
Acidophilic cytoplasm	light red – pink
Polychromatic cytoplasm	gray – violet
Acidophilic granules	orange
Neutrophilic granules	brown – pink
Basophilic granules	dark violet
Azurophilic granules	purple – violet



**MGG Quick Stain** 

PRODUCT AND APPLICATION		CODE	
MGG Quick Stain		04-090805	
Minimum number of tests that can be performed	100		
Completion time	20 seconds		
Shelf life	2 years		
Storage conditions	15-25°C		

## Application

Rapid method for differential staining of formed blood elements and other air-dried cellular smears.

Not required

#### Method

• 1) Dry the smear in air.

Additional equipment

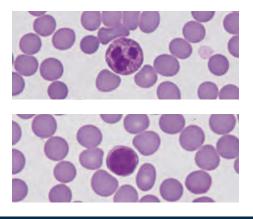
- 2) Immerse the slide 5 times for 1 second in solution A. After each immersion, wait a moment for the excess liquid to drain off.
- 3) Immerse the slide 5 times for 1 second in solution B. After each immersion, wait a moment for the excess liquid to drain off.
- 4) Immerse the slide 3-5 times for 1 second in the solution C. After each immersion, wait a moment for the excess liquid to drain off.
- 5) Wash in tap water.
- 6) Dry in the air (do not use heat sources, ovens or plates).

PRODUCT AND APPLICATION	PACK	CODE
MGG Quick Stain	0,5 I - 2 bottles of reagent A + 1 bottle of reagent B + 1 bottle of reagent C	04-090805M

#### Result

The colors and details are superimposable on those of May Grunwald Giemsa standard staining



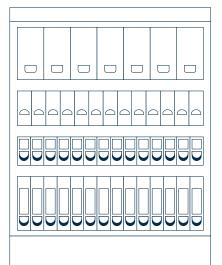


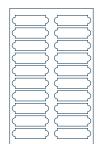




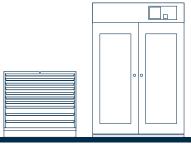








Storage





## **High-capacity filing cabinets**

Painted steel filing cabinets with blue epoxy powder coating, electrostatically applied, without solvents for environmental integrity, resistant to common chemical aggression. Each drawer is mounted on sliding guides, thus providing access to the entire surface area.

Its interior is equipped with plastic trays (code 03-C28N) for more effective archiving of both slides and embedded blocks.

They are equipped with central locking, available in three versions:

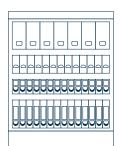
- Lock with security cylinder, including anti-tip system (only possible to open one drawer at a time)
- Code lock: uses a numerical combination in place of a key
- Remote lock: electronic system with manual remote controls







PRODUCT	CAPACITY (nr. DISHES)	DRAWERS	DIMENSIONS (1023x725xh)		CODE
For slides	67200 (140)	5	700 mm	150	03-V77000B
	94080 (196)	7	1000 mm	196	03-V109000B
	134400 (280)	10	1325 mm	267	03-V155000B
	147840 (308)	11	1450 mm	292	03-V171000B
	161280 (336)	12	1625 mm	300	03-V186500B
For cassettes	26880 (336)	12	1000 mm	274	03-B34000B
	31360 (392)	14	1150 mm	316	03-B39650B
	35840 (448)	16	1325 mm	351	03-B45320B
	40320 (504)	18	1450 mm	402	03-B51000B
	44800 (560)	20	1625 mm	447	03-B56600B
For slides and cassettes	80640 S	14	1450 mm	338	03-V92B23B
	17920 C	(S6 and C8)			
	80640 S	16	1550 mm	400	03-V80B22B
	22400 C	(S6 and C10)			





## Storage



## Wheeled filing cabinets

Archives for slides or blocks equipped with wheels and handles for transport. The top of the filing cabinet is equipped with a blue grooved mat with containment lip so that trays of slides can be placed on it without slipping and falling in transit. There are also three different types of lock to choose from for these filing cabinets:

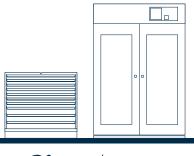
- Key lock
- Code lock
- Remote lock



PRODUCT	CAPACITY	DRAVERS	DISHES	DIMENSIONS MM	WEIGHT KG (EMPTY)	CODE
For cassettes	11520	12	144	564x725x1000	188	03-B13440B
For slides	51840	9	108	564x725x1000	159	03-V60480B
For slides and	34560 slides	10 (6 for slides and	120	564x725x1000	168	03-V40B4B
cassettes	3840 cassettes	4 for cassettes)				

## Accessories for high-capacity filing cabinets

PRODUCT	CODE
Tray for slides and embedded blocks	03-C28N
Tray slides and cassettes for large samples	03-C28S
Steel base for filing cabinets, designed for use with pallet trucks	03-90320120
Separation spring - 4 pcs	03-5000-MDL
Key lock	03-820.002
Code lock	03-820.011



# Bio - Optica

## **Modular filing cabinets**

## **Histoslide - Histoblock**

Modular systems for filing slides and paraffinembedded preparations.

The Histoslide modules (for slides) and Histoblock modules (for cassettes) are made entirely of white enameled metal.

They consist of sliding drawers fitted on guides. Each block of Histoslide/Histoblock modules requires completion with a base and top.

The special 7-drawer version for large slides or super mega cassettes retains the same modular form.

The spacer spring is recommended for keeping the slides in a vertical position with the correct spacing between them.

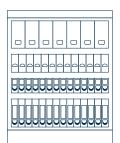






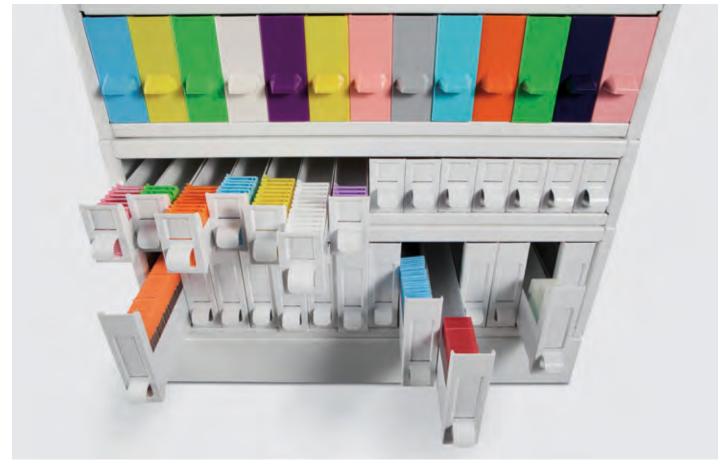
#### Characteristics

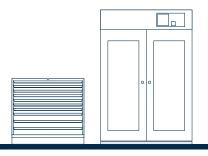
Dimensions (W x D x H):	Histoslide - 490 x 490 x 140 mm
	Histoblock - 490 x 490 x 90 mm
Weight per module:	13 kg empty, approximately 20 kg with cassettes,
	approximately 40 kg with slides
Capacity per module: Histoslide - up to 5000 slides	
	Histoblock - up to 860 cassettes or 540 rings
Recommended stacking of modules:	Up to 10 Histoslide
	15 Histoblock
Number of drawers per module:	14
PRODUCT	CODE
Histoslide for slides	03-5000-14
Histoblock for cassettes	03-B900
Histoslide with 7 drawers for large samples	s 03-7000
Base	03-5000-ВА
Тор	03-5000-CO
Spacer spring	03-5000-MD



## Storage

PRODUCT		CODE	Colorteca
Base		03-5000-BA	Modular system for filing slides and paraffin-
Тор		03-5000-C0	embedded preparations.
Metal structure		03-COLOR13	The plastic drawers are specifically designed
Plastic drawer	white with dividers for slides	03-CA7100S	to contain both slides and embedded
	orange with dividers for slides	03-CA7110S	preparations.
	light blue with dividers for slides	03-CA7120S	The drawers are available in 8 different
	yellow with dividers for slides	03-CA7130S	colors, which can easily be associated with
	lilac with dividers for slides	03-CA7140S	the colors of the slides or blocks stored in
	pink with dividers for slides	03-CA7150S	them.
	green with dividers for slides	03-CA7160S	Each module consists of 13 drawers and
	gray with dividers for slides	03-CA7180S	each drawer can contain approximately 330
	white	03-CA7100	slides or 48 blocks or 24 rings.
	orange	03-CA7110	The external dimensions of each module are
	blue	03-CA7120	the same as those of the Histoslide modules
	yellow	03-CA7130	(code 03- 5000-14) and Histoblock modules
	lilac	03-CA7140	(code 03- B900); this means you can stack
	pink	03-CA7150	Colorteca on top of an existing filing cabinet.
	green	03-CA7160	
	gray	03-CA7180	





Bio - Optica



#### **Bio Block**

Modular plastic 8-drawer filing cabinet for paraffin-embedded preparations (cassettes or rings). Each drawer has seven compartments. The total capacity of one Bio Block is approximately 2,250 cassettes. Bio Block is outstandingly modular thanks to its handy fastening system, which makes it possible to add modules both vertically and horizontally.

240x300x400 mm	1 pc	03 3000	
240x300x400 mm	1 pc.	03-3000	

## **Cartoglass - Cartoblock**

Modular systems in strong, lightweight cardboard, which are easy to transport even when completely filled.

The Cartoglass modules (for slides) and Cartobloc modules (for cassettes) are equipped with internal partitions, also made of cardboard, so as to create 36 compartments in the Cartoglass module and 16 compartments in the Cartobloc module.

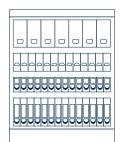
PRODUCT	CAPACITY PER MODULE	DIMENSIONS	PACK	CODE
Cartoglass Cartoblock	3000 slides 320 cassettes	290x400x80 mm 290x400x45 mm	10 pcs. 10 pcs.	03-4015-BA 03-4010-BC
Cover	220 rings	290x400x15 mm	10 pcs.	03-4020-C0



#### **Plastic slide boxes**

Stackable, made of shockproof material. Supplied complete with record form for the classification of preparations.

25 slides 98x83x38 mm 1 pc. 44-13071	
50 slides 230x97x35 mm 1 pc. 44-13072	
100 slides 230x180x35 mm 1 pc. 44-13073	





## Storage

## **Cardboard slide trays**

Trays for classifying and filing standard size slides (25x75 mm or 26x76 mm).

PRODUCT	PACK	DIMENSIONS	CODE
	4	100.01	~~~~~
2 slides with lid	1 pc.	102x94 mm	09-0002
6 slides with lid	1 pc.	213x102 mm	09-0006
10 slides with lid	1 pc.	342x102 mm	09-0010
20 slides	1 pc.	342x205 mm	09-0000
20 slides with dividers	1 pc.	342x205 mm	09-0020
20 slides with lid	1 pc.	342x205 mm	09-0001
20 slides with dividers and lid	1 pc.	342x205 mm	09-0023



## **Plastic slide trays**

Trays for classifying and filing standard size slides (25x75 mm or 26x76 mm).

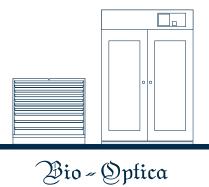
PRODUCT	PACK	DIMENSIONS	CODE
20 slides	20 pcs.	190x340 mm	44-13081
40 slides	10 pcs.	395x340 mm	44-13082



#### Slide mailers

Made of shock-proof plastic.

PRODUCT	PACK	DIMENSIONS	CODE
With press cap x5 slides	50 pcs.	28x82x16 mm	09-000530
With screw cap x5-10 slides	10 pcs.	ø 40xh 90 mm	44-13061
Snap-on x1 slide	500 pcs.	50x100x6 mm	44-13031
Snap-on x2 slides	500 pcs.	73x85x6 mm	44-13041
Snap-on x3 slides	100 pcs.	100x84x6 mm	44-13051
	±00 poo:	Teeve ive initi	11 10001



## Safety cabinets with extractor system

Cabinets with extractor system, designed for storing histological samples preserved in formalin, or storing chemicals and solvents.

#### **Construction features**

- Electrogalvanized steel structure
- Three tray-type shelves, height-adjustable
- Leaf doors made of safety glass

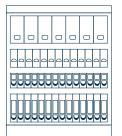
## Extractor system

- Non-sparking extractor
- Alumina pellet filters for formaldehyde
   Manifold for connection to central
- extractor systems
   Control panel with soft-touch keypad
- Control panel with soft-touch keypad for setting the desired operating parameters





PRODUCT	EXTERNAL DIMENSIONS	INTERNAL DIMENSIONS	CODE
Chemical Cabinet Formalin Cabinet	1200 x 550 x 1900 mm 1200 x 550 x 1900 mm		50-120-603 50-120-604
ACCESSORIES			CODE
Additional shelf			50-600-062
Formalin filter			50-F001
Solvent filter			50-F002
HEPA filter			50-F006
UV Lamp			50-600-051





Storage



## Contents

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AlcoolPath 95	32
Alkaline phosphatase	111
Amylase	63
ATPase	109
AUS240 stainer	46
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## В

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Bench Tech	49
Benchtop fume hood	49
Bielschowsky	65
Bio-Agar	120
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Biodec R	23
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Bio Marking Dyes	20
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Biopsy pads	27
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## С

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CVR909 coverslipper	47
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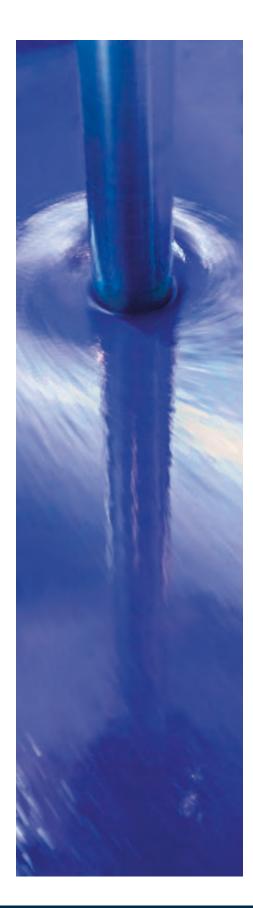
#### R

Rapid Frozen Sections

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120				
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Scott's water Screen-printed multi-purpose containers Silver impregnation Sirius Red Slide adapter for large samples Slide boxes Slide envelope Slide Master Slide trays Stainer automatic Succinate Dehydrogenase Sudan III Sudan Black	116 22 77 96 52 130 131 51 131 46 112 115 115
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